

REFERENCES

- Ahmed, H. M., Ramadhani, A. M., Erwa, I. Y., Ishag, O. A. O., & Saeed, M. B. (2020). Phytochemical screening, chemical composition and antimicrobial activity of cinnamon verum bark. *International Research Journal of Pure and Applied Chemistry*, 21(11), 36-43.
- Alara, O. R., Abdurahman, N. H., & Ukaegbu, C. I. (2021). Extraction of phenolic compounds: A review. *Current Research in Food Science*, 4, 200-214.
- Alara, O. R., Abdurahman, N. H., Ukaegbu, C. I., & Azhari, N. H. (2018). Vernonia cinerea leaves as the source of phenolic compounds, antioxidants, and anti-diabetic activity using microwave-assisted extraction technique. *Industrial Crops and Products*, 122, 533-544.
- Altemimi, A., Lakhssassi, N., Baharlouei, A., Watson, D. G., & Lightfoot, D. A. (2017). Phytochemicals: Extraction, isolation, and identification of bioactive compounds from plant extracts. *Plants*, 6(4), 42.
- Analuddin, K., Septiana, A., Nasaruddin, Sabilu, Y., & Sharma, S. (2019). Mangrove fruit bioprospecting: nutritional and antioxidant potential as a food source for coastal communities in the Rawa Aopa Watumohai National Park, Southeast Sulawesi, Indonesia. *International Journal of Fruit Science*, 19(4), 423-436.
- Annegowda, H. V., Gooi, T. S., Awang, S. H. H., Alias, N. A., & Mordi, M. N. (2012). Evaluation of Analgesic and Antioxidant Potency of Various Extracts of Cinnamomum iners Bark. *International journal of Pharmacology*, 8(3), 198-203.
- Aswar, A., Malik, A., Hamidu, L., & Najib, A. (2021). Determination of Total Phenolic Content of The Stem Bark Extract of Nyirih (*Xylocarpus granatum* J. Koeing) Using UV-Vis Spectrophotometry Method. *Jurnal Fitofarmaka Indonesia*, 8(3), 13-18.
- Awouafack, M. D., Tane, P., Kuete, V., & Eloff, J. N. (2013). Sesquiterpenes from the medicinal plants of Africa. In *Medicinal plant research in Africa* (pp. 33-103). Elsevier.
- Ayad, R., & Akkal, S. (2019). Phytochemistry and biological activities of algerian *Centaurea* and related genera. In *Studies in Natural Products Chemistry* (Vol. 63, pp. 357-414). Elsevier.
- Babbar, N., Oberoi, H. S., Sandhu, S. K., & Bhargav, V. K. (2014). Influence of different solvents in extraction of phenolic compounds from vegetable residues and their evaluation as natural sources of antioxidants. *Journal of food science and technology*, 51(10), 2568-2575.
- Barij, N., Čermák, J., & Stokes, A. (2011). Azimuthal variations in xylem structure and water relations in cork oak (*Quercus suber*). *Iawa Journal*, 32(1), 25-40.
- Beattie, A. J., Hay, M., Magnusson, B., de Nys, R., Smeathers, J., & Vincent, J. F. (2011). Ecology and bioprospecting. *Austral ecology*, 36(3), 341-356.

- Belay, R., & Makonnen, E. (2020). Anti-inflammatory activities of ethanol leaves extract and solvent fractions of *Zehneria scabra* (Cucurbitaceae) in rodents. *Asian Journal of Natural Product Biochemistry*, 18(1).
- Bele, A. A., & Khale, A. (2011). An overview on thin layer chromatography. *International journal of pharmaceutical sciences and research*, 2(2), 256.
- Bernard-Savary, P., & Poole, C. F. (2015). Instrument platforms for thin-layer chromatography. *Journal of Chromatography A*, 1421, 184-202.
- Berthier, E., Warrick, J., Yu, H., & Beebe, D. J. (2008). Managing evaporation for more robust microscale assays Part 1. Volume loss in high throughput assays. *Lab on a Chip*, 8(6), 852-859.
- Betz, J. M., Brown, P. N., & Roman, M. C. (2011). Accuracy, precision, and reliability of chemical measurements in natural products research. *Fitoterapia*, 82(1), 44-52.
- Bharudin, M. A., Zakaria, S., & Chia, C. H. (2013, November). Condensed tannins from acacia mangium bark: Characterization by spot tests and FTIR. In *AIP Conference Proceedings* (Vol. 1571, No. 1, pp. 153-157). American Institute of Physics.
- Blainski, A., Lopes, G. C., & De Mello, J. C. P. (2013). Application and analysis of the folin ciocalteu method for the determination of the total phenolic content from *Limonium brasiliense* L. *Molecules*, 18(6), 6852-6865.
- Boeing, J. S., Barizão, É. O., Montanher, P. F., de Cinque Almeida, V., & Visentainer, J. V. (2014). Evaluation of solvent effect on the extraction of phenolic compounds and antioxidant capacities from the berries: application of principal component analysis. *Chemistry central journal*, 8(1), 1-9.
- Botelho, G., Canas, S., & Lameiras, J. (2017). Development of phenolic compounds encapsulation techniques as a major challenge for food industry and for health and nutrition fields. In *Nutrient Delivery* (pp. 535-586). Academic Press.
- Boysen, R. I., & Hearn, M. T. W. (2010). High performance liquid chromatographic separation methods. In *Comprehensive Natural Products II: Chemistry and Biology* (pp. 5-49). Elsevier.
- Cai, L. (2014). Thin layer chromatography. *Current Protocols Essential Laboratory Techniques*, 8(1), 6-3.
- Carmona-Hernandez, J. C., Taborda-Ocampo, G., & González-Correa, C. H. (2021). Folin-ciocalteu reaction alternatives for higher polyphenol quantitation in Colombian passion fruits. *International Journal of Food Science*, 2021.
- Cassiday, L. (2014). Emulsions: making oil and water mix. *International News on Fats, Oils and Related Materials*, 25(4), 200-208.

- Chandrasekara, A., & Josheph Kumar, T. (2016). Roots and tuber crops as functional foods: a review on phytochemical constituents and their potential health benefits. *International journal of food science*, 2016.
- Charles, D. J. (2012). Cinnamon. In *Antioxidant Properties of Spices, Herbs and Other Sources* (pp. 231-243). Springer, New York, NY.
- Che Sulaiman, I. S., Basri, M., Fard Masoumi, H. R., Chee, W. J., Ashari, S. E., & Ismail, M. (2017). Effects of temperature, time, and solvent ratio on the extraction of phenolic compounds and the anti-radical activity of *Clinacanthus nutans* Lindau leaves by response surface methodology. *Chemistry Central Journal*, 11(1), 1-11.
- Chung, C., Sher, A., Rousset, P., Decker, E. A., & McClements, D. J. (2017). Formulation of food emulsions using natural emulsifiers: Utilization of quillaja saponin and soy lecithin to fabricate liquid coffee whiteners. *Journal of Food Engineering*, 209, 1-11.
- Co, M., Fagerlund, A., Engman, L., Sunnerheim, K., Sjöberg, P. J., & Turner, C. (2012). Extraction of antioxidants from spruce (*Picea abies*) bark using eco-friendly solvents. *Phytochemical analysis*, 23(1), 1-11.
- Ćujić, N., Šavikin, K., Janković, T., Pljevljakušić, D., Zdunić, G., & Ibrić, S. (2016). Optimization of polyphenols extraction from dried chokeberry using maceration as traditional technique. *Food chemistry*, 194, 135-142.
- Cushnie, T. T., Cushnie, B., Echeverría, J., Fowsantear, W., Thammawat, S., Dodgson, J. L., ... & Clow, S. M. (2020). Bioprospecting for antibacterial drugs: A multidisciplinary perspective on natural product source material, bioassay selection and avoidable pitfalls. *Pharmaceutical Research*, 37(7), 1-24.
- Dey, P., Kundu, A., Kumar, A., Gupta, M., Lee, B. M., Bhakta, T., ... & Kim, H. S. (2020). Analysis of alkaloids (indole alkaloids, isoquinoline alkaloids, tropane alkaloids). In *Recent advances in natural products analysis* (pp. 505-567). Elsevier.
- Diouf, P. N., Stevanovic, T., & Cloutier, A. (2009). Study on chemical composition, antioxidant and anti-inflammatory activities of hot water extract from *Picea mariana* bark and its proanthocyanidin-rich fractions. *Food Chemistry*, 113(4), 897-902.
- Do, Q. D., Angkawijaya, A. E., Tran-Nguyen, P. L., Huynh, L. H., Soetaredjo, F. E., Ismadji, S., & Ju, Y. H. (2014). Effect of extraction solvent on total phenol content, total flavonoid content, and antioxidant activity of *Limnophila aromatica*. *Journal of food and drug analysis*, 22(3), 296-302.

- Dogan, K., Akman, P. K., & Tornuk, F. (2019). Tree Barks as Potential Sources of Value-added Components for the Food Industry. *International Journal of Food Technology and Nutrition*, 2(3-4), 25-35.
- Donato-Capel, L., Garcia-Rodenas, C. L., Pouteau, E., Lehmann, U., Srichuwong, S., Erkner, A., ... & Sagalowicz, L. (2014). Technological means to modulate food digestion and physiological response. In *Food structures, digestion and health* (pp. 389-422). Academic Press.
- Duncan, S. E., & Chang, H. H. (2012). Implications of light energy on food quality and packaging selection. *Advances in food and nutrition research*, 67, 25-73.
- Ejikeme, C., Ezeonu, C. S., & Eboatu, A. N. (2014). Determination of Physical and Phytochemical Constituents of some Tropical Timbers Indigenous to nigerdelta area of nigeria. *European Scientific Journal*, 10(18), 247-270.
- El Hajaji, H., Lachkar, N., Alaoui, K., Cherrah, Y., Farah, A., Ennabili, A., ... & Lachkar, M. (2011). Antioxidant activity, phytochemical screening, and total phenolic content of extracts from three genders of carob tree barks growing in Morocco. *Arabian Journal of Chemistry*, 4(3), 321-324.
- Ezeonu, C. S., & Ejikeme, C. M. (2016). Qualitative and quantitative determination of phytochemical contents of indigenous Nigerian softwoods. *New Journal of Science*, 2016.
- Ezez, D., & Tefera, M. (2021). Effects of solvents on total phenolic content and antioxidant activity of ginger extracts. *Journal of Chemistry*, 2021.
- Fedorov, V. S., & Ryazanova, T. V. (2021). Bark of Siberian conifers: Composition, use, and processing to extract tannin. *Forests*, 12(8), 1043.
- Felhi, S., Daoud, A., Hajlaoui, H., Mnafigui, K., Gharsallah, N., & Kadri, A. (2017). Solvent extraction effects on phytochemical constituents profiles, antioxidant and antimicrobial activities and functional group analysis of *Ecballium elaterium* seeds and peels fruits. *Food Science and Technology*, 37, 483-492.
- Feng, S., Cheng, S., Yuan, Z., Leitch, M., & Xu, C. C. (2013). Valorization of bark for chemicals and materials: A review. *Renewable and Sustainable Energy Reviews*, 26, 560-578.
- Ferdes, M. (2018). Antimicrobial compounds from plants. *Fighting Antimicrobial Resistance. IAPC-OBP, Zagreb*, 243-271.
- Ferreira, J. P., Miranda, I., Sousa, V. B., & Pereira, H. (2018). Chemical composition of barks from *Quercus faginea* trees and characterization of their lipophilic and polar extracts. *PLoS One*, 13(5), e0197135.
- Foo, L. Y., & Karchesy, J. J. (1989). Chemical nature of phlobaphenes. In *Chemistry and significance of condensed tannins* (pp. 109-118). Springer, Boston, MA.

- Gachelin, G., Garner, P., Ferroni, E., Tröhler, U., & Chalmers, I. (2017). Evaluating Cinchona bark and quinine for treating and preventing malaria. *Journal of the Royal Society of Medicine*, 110(1), 31-40.
- Galanakis, C. M. (Ed.). (2021). *Nutraceutical and functional food components: Effects of innovative processing techniques*. Academic Press.
- Gan, R. Y., Chan, C. L., Yang, Q. Q., Li, H. B., Zhang, D., Ge, Y. Y., ... & Corke, H. (2019). Bioactive compounds and beneficial functions of sprouted grains. In *Sprouted grains* (pp. 191-246). AACC International Press.
- Ghasemzadeh, A., Jaafar, H. Z., Bukhori, M. F. M., Rahmat, M. H., & Rahmat, A. (2018). Assessment and comparison of phytochemical constituents and biological activities of bitter bean (*Parkia speciosa* Hassk.) collected from different locations in Malaysia. *Chemistry central journal*, 12(1), 1-9.
- Ghatak, A., Nair, S., Vajpayee, A., Chaturvedi, P., Samant, S., Soley, K., ... & Desai, N. (2015). Evaluation of antioxidant activity, total phenolic content, total flavonoids, and LC-MS characterization of *Saraca asoca* (Roxb.) De. Wilde. *Int J Adv Res*, 3(5), 318-27.
- Giada, M. D. L. R. (2013). Food phenolic compounds: main classes, sources and their antioxidant power. *Oxidative stress and chronic degenerative diseases-A role for antioxidants*, 2013, 87-112.
- Gujjeti, R. P., & Mamidala, E. (2013a). Phytochemical analysis and TLC profile of *Madhuca indica* inner bark plant extract. *Inter J of Sci & Engin Res*, 4(10), 1507-1510.
- Gujjeti, R. P., & Mamidala, E. (2013b). Phytochemical screening and thin layer chromatographic studies of *Aerva lanata* root extract. *International journal of innovative Research in Science, Engineering and Technology*, 2(10), 5725-30.
- Haminiuk, C. W. I., Plata-Oviedo, M. S. V., de Mattos, G., Carpes, S. T., & Branco, I. G. (2014). Extraction and quantification of phenolic acids and flavonols from *Eugenia pyriformis* using different solvents. *Journal of Food Science and Technology*, 51(10), 2862-2866.
- Haryanto, A., Hidayat, W., Hasanudin, U., Iryani, D. A., Kim, S., Lee, S., & Yoo, J. (2021). Valorization of Indonesian wood wastes through pyrolysis: A review. *Energies*, 14(5), 1407.
- Hatami, T., Emami, S. A., Miraghaee, S. S., & Mojarrab, M. (2014). Total phenolic contents and antioxidant activities of different extracts and fractions from the aerial parts of *Artemisia biennis* Willd. *Iranian journal of pharmaceutical research: IJPR*, 13(2), 551.
- Hofmann, T., Visiné Rajczi, E., Bocz, B., Bocz, D., & Albert, L. (2020). Antioxidant Capacity and Tentative Identification of Polyphenolic Compounds of Cones of Selected Coniferous Species. *ACTA SILVATICA ET LIGNARIA HUNGARICA*, 16(2), 79-94.

- Houghton, P., & Raman, A. (2012). *Laboratory handbook for the fractionation of natural extracts*. Springer Science & Business Media.
- Huang, D. F., Xu, J. G., Liu, J. X., Zhang, H., & Hu, Q. P. (2014). Chemical constituents, antibacterial activity and mechanism of action of the essential oil from *Cinnamomum cassia* bark against four food-related bacteria. *Microbiology*, *83*(4), 357-365.
- Iravani, S., & Zolfaghari, B. (2014). Phytochemical analysis of *Pinus eldarica* bark. *Research in pharmaceutical sciences*, *9*(4), 243.
- Islam, E., Islam, R., Rahman, A. A., Alam, A. H. M., Khondkar, P., Rashid, M., & Parvin, S. (2013). Estimation of total phenol and in vitro antioxidant activity of *Albizia procera* leaves. *BMC research notes*, *6*(1), 1-7.
- Jadid, N., Hidayati, D., Hartanti, S. R., Arraniry, B. A., Rachman, R. Y., & Wikanta, W. (2017, June). Antioxidant activities of different solvent extracts of *Piper retrofractum* Vahl. using DPPH assay. In AIP conference proceedings (Vol. 1854, No. 1, p. 020019). AIP Publishing LLC.
- Jimenez-Garcia, S. N., Vazquez-Cruz, M. A., Garcia-Mier, L., Contreras-Medina, L. M., Guevara-González, R. G., Garcia-Trejo, J. F., & Feregrino-Perez, A. A. (2018). Phytochemical and Pharmacological Properties of Secondary Metabolites in Berries. *Therapeutic Foods*, 397-427.
- Johari, M. A., & Khong, H. Y. (2019). Total phenolic content and antioxidant and antibacterial activities of *Pereskia bleo*. *Advances in pharmacological sciences*, 2019.
- Kadum, H., Hamid, A. A., Abas, F., Ramli, N. S., Mohammed, A. K. S., Muhiadin, B. J., & Jaafar, A. H. (2019). Bioactive compounds responsible for antioxidant activity of different varieties of date (*Phoenix dactylifera* L.) elucidated by ¹H-NMR based metabolomics. *International Journal of Food Properties*, *22*(1), 462-476.
- Khaliq, H. A. (2016). Pharmacognostic, physiochemical, phytochemical and pharmacological studies on *Careya arborea* Roxb: a review. *J Phytopharmacol*, *5*(1), 27-34.
- Kim, Y. G., Lee, J. H., Kim, S. I., Baek, K. H., & Lee, J. (2015). Cinnamon bark oil and its components inhibit biofilm formation and toxin production. *International journal of food microbiology*, *195*, 30-39.
- Kregiel, D., Berlowska, J., Witonska, I., Antolak, H., Proestos, C., Babic, M., ... & Zhang, B. (2017). Saponin-based, biological-active surfactants from plants. *Application and characterization of surfactants*, *6*(1), 184-205.
- Kremer, D., Kosalec, I., Locatelli, M., Epifano, F., Genovese, S., Carlucci, G., & Končić, M. Z. (2012). Anthraquinone profiles, antioxidant and antimicrobial properties of *Frangula rupestris* (Scop.) Schur and *Frangula alnus* Mill. bark. *Food Chemistry*, *131*(4), 1174-1180.

- Krishnan, V., Ahmad, S., & Mahmood, M. (2015). Antioxidant potential in different parts and callus of *Gynura procumbens* and different parts of *Gynura bicolor*. *BioMed Research International*, 2015.
- Kumar, S., Jyotirmayee, K., & Sarangi, M. (2013). Thin layer chromatography: a tool of biotechnology for isolation of bioactive compounds from medicinal plants. *International Journal of Pharmaceutical Sciences Review and Research*, 18(1), 126-132.
- Lalhminghlu, K., & Jagetia, G. C. (2018). Evaluation of the free-radical scavenging and antioxidant activities of *Chilauni*, *Schima wallichii* Korth in vitro. *Future science OA*, 4(2), FSO272.
- Lamuela-Raventós, R. M. (2017). Folin-Ciocalteu method for the measurement of total phenolic content and antioxidant capacity. *Meas. Antioxid. Act. Capacit. Recent Trends Appl*, 107-115.
- Laura, A., Moreno-Escamilla, J. O., Rodrigo-García, J., & Alvarez-Parrilla, E. (2019). Phenolic compounds. In *Postharvest physiology and biochemistry of fruits and vegetables* (pp. 253-271). Woodhead Publishing.
- Lawes, M. J., Richards, A., Dathe, J., & Midgley, J. J. (2011). Bark thickness determines fire resistance of selected tree species from fire-prone tropical savanna in north Australia. *Plant Ecology*, 212(12), 2057-2069.
- Lawes, M. J., Midgley, J. J., & Clarke, P. J. (2013). Costs and benefits of relative bark thickness in relation to fire damage: a savanna/forest contrast. *Journal of Ecology*, 101(2), 517-524.
- Liang, N., & Kitts, D. D. (2014). Antioxidant property of coffee components: assessment of methods that define mechanisms of action. *Molecules*, 19(11), 19180-19208.
- Lin, D., Xiao, M., Zhao, J., Li, Z., Xing, B., Li, X., ... & Chen, S. (2016). An overview of plant phenolic compounds and their importance in human nutrition and management of type 2 diabetes. *Molecules*, 21(10), 1374.
- Littler, S. (2018). The importance and effect of sample size. Select Statistical Service.
- Loram-Lourenco, L., Farnese, F. D. S., Sousa, L. F. D., Alves, R. D. F. B., Andrade, M. C. P. D., Almeida, S. E. D. S., ... & Menezes-Silva, P. E. (2020). A structure shaped by fire, but also water: ecological consequences of the variability in bark properties across 31 species from the Brazilian Cerrado. *Frontiers in plant science*, 10, 1718.
- Macêdo, N. S., Silveira, Z. D. S., Bezerra, A. H., Costa, J. G. M. D., Coutinho, H. D. M., Romano, B., ... & da Silva, M. V. (2020). *Caesalpinia ferrea* C. Mart.(Fabaceae) phytochemistry, ethnobotany, and bioactivities: a review. *Molecules*, 25(17), 3831.
- Maisuthisakul, P., & Gordon, M. H. (2014). Characterization and storage stability of the extract of Thai mango (*Mangifera indica* Linn. Cultivar Chok-Anan) seed kernels. *Journal of food science and technology*, 51(8), 1453-1462.

- Makin, H. L., & Gower, D. B. (Eds.). (2010). *Steroid analysis*. Springer Science & Business Media.
- Malta, L. G., & Liu, R. H. (2014). Analyses of total phenolics, total flavonoids, and total antioxidant activities in foods and dietary supplements.
- Mármol, I., Quero, J., Jiménez-Moreno, N., Rodríguez-Yoldi, M. J., & Ancín-Azpilicueta, C. (2019). A systematic review of the potential uses of pine bark in food industry and health care. *Trends in Food Science & Technology*, *88*, 558-566.
- Mathew, S., Abraham, T. E., & Zakaria, Z. A. (2015). Reactivity of phenolic compounds towards free radicals under in vitro conditions. *Journal of food science and technology*, *52*(9), 5790-5798.
- Mihara, R., Barry, K. M., Mohammed, C. L., & Mitsunaga, T. (2005). Comparison of antifungal and antioxidant activities of *Acacia mangium* and *A. auriculiformis* heartwood extracts. *Journal of chemical ecology*, *31*(4), 789-804.
- Millum, J. (2010). How should the benefits of bioprospecting be shared?. *Hastings Center Report*, *40*(1), 24-33.
- Mondal, S. (2017). *Screening of bioactive compounds from Cucurbitaceae family edible plants of Bangladesh—Cucurbita pepo Linn.: A case study*. Anchor Academic Publishing.
- Nakbanpote, W., Ruttanakorn, M., Sukadeetad, K., Sakkayawong, N., & Damrianant, S. (2019). Effects of drying and extraction methods on phenolic compounds and in vitro assays of *Eclipta prostrata* Linn. leaf extracts. *Sci. Asia*, *45*, 127-137.
- Nariya, P. B., Bhalodia, N. R., Shukla, V. J., Acharya, R., & Nariya, M. B. (2013). In vitro evaluation of antioxidant activity of *Cordia dichotoma* (Forst f.) bark. *Ayu*, *34*(1), 124.
- Navid, M. H., Laszczyk-Lauer, M. N., Reichling, J., & Schnitzler, P. (2014). Pentacyclic triterpenes in birch bark extract inhibit early step of herpes simplex virus type 1 replication. *Phytomedicine*, *21*(11), 1273-1280.
- Nawaz, H., Shad, M. A., Rehman, N., Andaleeb, H., & Ullah, N. (2020). Effect of solvent polarity on extraction yield and antioxidant properties of phytochemicals from bean (*Phaseolus vulgaris*) seeds. *Brazilian Journal of Pharmaceutical Sciences*, *56*.
- Nebehaj, E., Albert, L., Visiné Rajczi, E., & Hofmann, T. (2019). Combined multi-assay evaluation of the antioxidant properties of tree bark. *ACTA SILVATICA ET LIGNARIA HUNGARICA: AN INTERNATIONAL JOURNAL IN FOREST, WOOD AND ENVIRONMENTAL SCIENCES*, *15*(2), 85-97.
- Nesfu, N. Z. M. (2020). The studies on *Momordica charantia* fruits and the synthesis of pyrazolyl substituted benzylidene indanone derivatives as dengue virus type-2 NS2B/NS3 protease inhibitor (Doctoral dissertation, Université de Lorraine; Universiti Sains Malaysia (Malaisie)).
- Neumann, M., & Lawes, M. J. (2021). Quantifying carbon in tree bark: The importance of bark morphology and tree size. *Methods in Ecology and Evolution*, *12*(4), 646-654.

- Ngo, T. V., Scarlett, C. J., Bowyer, M. C., Ngo, P. D., & Vuong, Q. V. (2017). Impact of different extraction solvents on bioactive compounds and antioxidant capacity from the root of *Salacia chinensis* L. *Journal of Food Quality*, 2017.
- Noral, A., Wilapanggal, A., & Novianti, T. (2018). Antioxidant activity, antibacterial activity, water content, and ash content in Baduy honey. *Bioscience*, 2(1), 38-44.
- Nur-Hadirah, K., Arifullah, M., Nazahatul, A. A., Klaiklay, S., Chumkaew, P., Norhazlini, M. Z., & Zulhazman, H. (2021, May). Total phenolic content and antioxidant activity of an edible Aroid, *Colocasia esculenta* (L.) Schott. In *IOP Conference Series: Earth and Environmental Science* (Vol. 756, No. 1, p. 012044). IOP Publishing.
- Olejar, K. J., Fedrizzi, B., & Kilmartin, P. A. (2015). Influence of harvesting technique and maceration process on aroma and phenolic attributes of Sauvignon blanc wine. *Food chemistry*, 183, 181-189.
- Ostroukhova, L., Fedorova, T., Onuchina, N., Levchuk, A. and Babkin, V., (2018). Determination of Quantitative Content of Extractive Substances from the Wood, Roots, and Bark of Coniferous Species in Siberia: Larch (*Larix Sibirica* L.), Scotch Pine (*Pinus Sylvestris* L.), Fir (*Abies Sibirica* L.), Spruce (*Picea Obovata* L.) AND Cedar (*Pinus Sibirica* Du Tour). *chemistry of plant raw material*, (4), pp.185-195.
- Oyemitan, I. A. (2017). *African Medicinal Spices of Genus Piper. Medicinal Spices and Vegetables from Africa*, 581–597. doi:10.1016/b978-0-12-809286-6.00027-3
- Palma, G., Freer, J., & Baeza, J. (2003). Removal of metal ions by modified *Pinus radiata* bark and tannins from water solutions. *Water Research*, 37(20), 4974-4980.
- Panche, A. N., Diwan, A. D., & Chandra, S. R. (2016). Flavonoids: an overview. *Journal of nutritional science*, 5.
- Pandey, A., & Tripathi, S. (2014). Concept of standardization, extraction and pre phytochemical screening strategies for herbal drug. *Journal of Pharmacognosy and Phytochemistry*, 2(5).
- Parbuntari, H., Prestica, Y., Gunawan, R., Nurman, M. N., & Adella, F. (2018). Preliminary phytochemical screening (qualitative analysis) of cacao leaves (*Theobroma cacao* L.). *Eksakta: Berkala Ilmiah Bidang MIPA (E-ISSN: 2549-7464)*, 19(2), 40-45.
- Parham, S., Kharazi, A. Z., Bakhsheshi-Rad, H. R., Nur, H., Ismail, A. F., Sharif, S., ... & Berto, F. (2020). Antioxidant, antimicrobial and antiviral properties of herbal materials. *Antioxidants*, 9(12), 1309.
- Pasztory, Z., Mohácsiné, I. R., Gorbacheva, G., & Böröcsök, Z. (2016). The utilization of tree bark. *BioResources*, 11(3), 7859-7888.

- Patel, S. (2016). Plant-derived cardiac glycosides: Role in heart ailments and cancer management. *Biomedicine & Pharmacotherapy*, *84*, 1036-1041.
- Ponomarenko, J., Trouillas, P., Martin, N., Dizhbite, T., Krasilnikova, J., & Telysheva, G. (2014). Elucidation of antioxidant properties of wood bark derived saturated diarylheptanoids: A comprehensive (DFT-supported) understanding. *Phytochemistry*, *103*, 178-187.
- Prakash, V. E. D. (2017). Terpenoids as source of anti-inflammatory compounds. *Asian Journal of Pharmaceutical and Clinical Research*, *10*(3), 68-76.
- Puspitasari, L., Swastini, D. A., & Arisanti, C. I. A. (2013). Skrining fitokimia ekstrak etanol 95% kulit buah manggis (*Garcinia mangostana* L.). *Jurnal Farmasi Udayana*, *2*(3), 1-4.
- Radhia, A., Hanen, N., Abdelkarim, B. A., & Mohamed, N. (2018). Phytochemical screening, antioxidant and antimicrobial activities of *Erodium glaucophyllum* (L.) L'Hérit. *Journal of Biomedical Sciences*, *7*(4).
- Rahayuningsih, E., Mindaryani, A., Adriyanti, D. T., Parthasiwi, L. D., Adina, H. P., & Dyah, A. E. (2020, April). Conceptual Design of a Process Plant for the Production of Natural Dye from Merbau (*Intsia bijuga*) Bark. In IOP Conference Series: Materials Science and Engineering (Vol. 778, No. 1, p. 012045). IOP Publishing.
- Rahman, M. M., Islam, M. B., Biswas, M., & Alam, A. K. (2015). In vitro antioxidant and free radical scavenging activity of different parts of *Tabebuia pallida* growing in Bangladesh. *BMC research notes*, *8*(1), 1-9.
- Rani, P., Yu, X., Liu, H., Li, K., He, Y., Tian, H., & Kumar, R. (2021). Material, antibacterial and anticancer properties of natural polyphenols incorporated soy protein isolate: A review. *European Polymer Journal*, *152*, 110494.
- Rao, P. V., & Gan, S. H. (2014). Cinnamon: a multifaceted medicinal plant. *Evidence-Based Complementary and Alternative Medicine*, *2014*.
- Rivero-Cruz, J. F., Granados-Pineda, J., Pedraza-Chaverri, J., Pérez-Rojas, J. M., Kumar-Passari, A., Diaz-Ruiz, G., & Rivero-Cruz, B. E. (2020). Phytochemical constituents, antioxidant, cytotoxic, and antimicrobial activities of the ethanolic extract of Mexican brown propolis. *Antioxidants*, *9*(1), 70.
- Robinson, D. F. (2012). Biopiracy and the innovations of indigenous peoples and local communities. *Indigenous People's Innovation: Intellectual Property Pathways to Development*, 77-78.
- Rohman, A. (2014). Rice bran oil's role in health and cooking. In *Wheat and Rice in Disease Prevention and Health* (pp. 481-490). Academic Press.

- Rosell, J. A., Olson, M. E., Anfodillo, T., & Martínez-Méndez, N. (2017). Exploring the bark thickness–stem diameter relationship: clues from lianas, successive cambia, monocots and gymnosperms. *New Phytologist*, *215*(2), 569-581.
- Saeed, N., Khan, M. R., & Shabbir, M. (2012). Antioxidant activity, total phenolic and total flavonoid contents of whole plant extracts *Torilis leptophylla* L. *BMC complementary and alternative medicine*, *12*(1), 1-12.
- Salem, M. Z. M., Elansary, H. O., Elkelish, A. A., Zeidler, A., Ali, H. M., Mervat, E. H., & Yessoufou, K. (2016). In vitro bioactivity and antimicrobial activity of *Picea abies* and *Larix decidua* wood and bark extracts. *BioResources*, *11*(4), 9421-9437.
- Salem, N. M., & Awwad, A. M. (2014). Biosorption of Ni (II) from electroplating wastewater by modified (*Eriobotrya japonica*) loquat bark. *Journal of Saudi Chemical Society*, *18*(5), 379-386.
- Santos, M. C., & Gonçalves, É. C. (2016). Effect of different extracting solvents on antioxidant activity and phenolic compounds of a fruit and vegetable residue flour. *Scientia agropecuaria*, *7*(1), 7-14.
- Santos, S. A., Pinto, P. C., Silvestre, A. J., & Neto, C. P. (2010). Chemical composition and antioxidant activity of phenolic extracts of cork from *Quercus suber* L. *Industrial Crops and Products*, *31*(3), 521-526.
- Saxena, J., Baunthiyal, M., & Ravi, I. (2015). *Laboratory Manual of Microbiology, Biochemistry and Molecular Biology*. Scientific Publishers.
- Schafer, J. L., Breslow, B. P., Hohmann, M. G., & Hoffmann, W. A. (2015). Relative bark thickness is correlated with tree species distributions along a fire frequency gradient. *Fire Ecology*, *11*(1), 74-87.
- Seidel, V. (2012). Initial and bulk extraction of natural products isolation. *Natural products isolation*, 27-41.
- Sembing, E. N., Elya, B., & Sauriasari, R. (2018). Phytochemical screening, total flavonoid and total phenolic content and antioxidant activity of different parts of *Caesalpinia bonduc* (L.) Roxb. *Pharmacognosy journal*, *10*(1).
- Sen, A., Zhianski, M., Glushkova, M., Petkova, K., Ferreira, J., & Pereira, H. (2016). Chemical composition and cellular structure of corks from *Quercus suber* trees planted in Bulgaria and Turkey. *Wood Science and Technology*, *50*(6), 1261-1276.
- Shantabi, L., & Jagetia, G. C. (2015). Phytochemical profiling of Kam-sabut, *Croton caudatus* Geiseler. *Res Rev J Bot Sci*, *4*, 5-14.

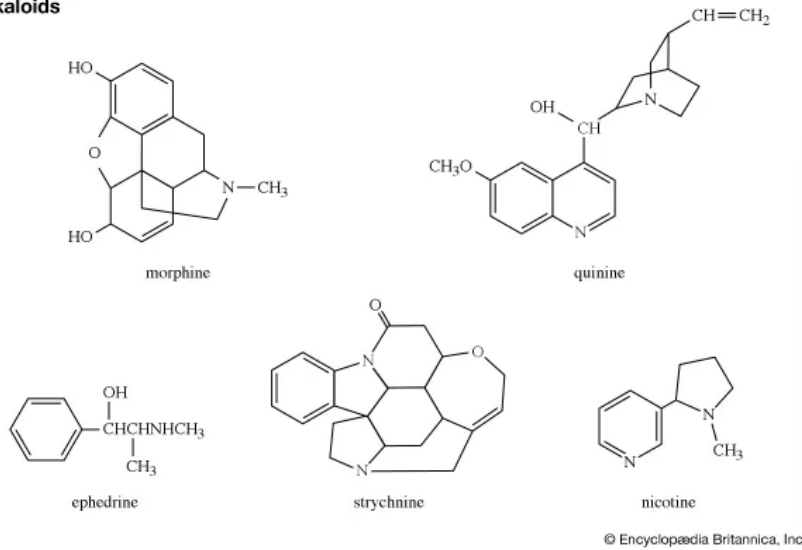
- Sharma, K., Kumar, V., Kaur, J., Tanwar, B., Goyal, A., Sharma, R., ... & Kumar, A. (2021). Health effects, sources, utilization and safety of tannins: A critical review. *Toxin Reviews*, *40*(4), 432-444.
- Shi, J., Arunasalam, K., Yeung, D., Kakuda, Y., Mittal, G., & Jiang, Y. (2004). Saponins from edible legumes: chemistry, processing, and health benefits. *Journal of medicinal food*, *7*(1), 67-78.
- Shi, Q. I. U., Hui, S. U. N., Zhang, A. H., Hong-Ying, X. U., Guang-Li, Y. A. N., Ying, H. A. N., & Xi-Jun, W. A. N. G. (2014). Natural alkaloids: basic aspects, biological roles, and future perspectives. *Chinese Journal of Natural Medicines*, *12*(6), 401-406.
- Sholichah, A. R. (2017, November). Phytochemical screening and antioxidant activity of ethanolic extract and ethyl acetate fraction from basil leaf (*Ocimum basilicum* L.) by DPPH radical scavenging method. In *IOP Conference Series: Materials Science and Engineering* (Vol. 259, No. 1, p. 012008). IOP Publishing.
- Škrovánková, S., Mišurcová, L., & Machů, L. (2012). Antioxidant activity and protecting health effects of common medicinal plants. *Advances in food and nutrition research*, *67*, 75-139.
- Somarróm, N., & Areekul, V. (2020). Extraction Optimization of Total Phenolic Content and Antioxidant Activity from Teaw (*Cratoxylum Formosum*) by Central Composite Design.
- Stängle, S. M., Sauter, U. H., & Dormann, C. F. (2017). Comparison of models for estimating bark thickness of *Picea abies* in southwest Germany: the role of tree, stand, and environmental factors. *Annals of forest science*, *74*(1), 1-10.
- Szwajkowska-Michałek, L., Przybylska-Balcerek, A., Rogoziński, T., & Stuper-Szablewska, K. (2020). Phenolic Compounds in Trees and Shrubs of Central Europe. *Applied Sciences*, *10*(19), 6907.
- Taib, M., Rezzak, Y., Bouyazza, L., & Lyoussi, B. (2020). Medicinal uses, phytochemistry, and pharmacological activities of *Quercus* Species. *Evidence-Based Complementary and Alternative Medicine*, 2020.
- Telichowska, A., Kobus-Cisowska, J., & Szulc, P. (2020). Phytopharmacological possibilities of bird cherry *Prunus padus* L. and *Prunus serotina* L. species and their bioactive phytochemicals. *Nutrients*, *12*(7), 1966.
- Telichowska, A., Kobus-Cisowska, J., Szulc, P., Wilk, R., Szwajgier, D., & Szymanowska, D. (2021). *Prunus padus* L. bark as a functional promoting component in functional herbal infusions—cyclooxygenase-2 inhibitory, antioxidant, and antimicrobial effects. *Open Chemistry*, *19*(1), 1052-1061.
- Thamizhiniyan, V., Young-Woong, C., & Young-Kyoon, K. (2015). The cytotoxic nature of *Acanthopanax sessiliflorus* stem bark extracts in human breast cancer cells. *Saudi journal of biological sciences*, *22*(6), 752-759.

- Thouri, A., Chahdoura, H., El Arem, A., Omri Hichri, A., Ben Hassin, R., & Achour, L. (2017). Effect of solvents extraction on phytochemical components and biological activities of Tunisian date seeds (var. Korkobbi and Arechti). *BMC complementary and alternative medicine*, 17(1), 1-10.
- Tong, W. (2013). Biotransformation of terpenoids and steroids. *Natural Products*, Springer, 2733-2759.
- Tungmunnithum, D., Thongboonyou, A., Pholboon, A., & Yangsabai, A. (2018). Flavonoids and other phenolic compounds from medicinal plants for pharmaceutical and medical aspects: An overview. *Medicines*, 5(3), 93.
- Vane, C. H., Drage, T. C., & Snape, C. E. (2006). Bark decay by the white-rot fungus *Lentinula edodes*: Polysaccharide loss, lignin resistance and the unmasking of suberin. *International Biodeterioration & Biodegradation*, 57(1), 14-23.
- Vecera, M. (2012). *Detection and identification of organic compounds*. Springer Science & Business Media.
- Venkatesan, T., Choi, Y. W., & Kim, Y. K. (2019). Impact of different extraction solvents on phenolic content and antioxidant potential of *Pinus densiflora* bark extract. *BioMed research international*, 2019.
- Verpoorte, R. (2021). *Eutectic Solvents and Stress in Plants*. Academic Press.
- Wadood, A., Ghufuran, M., Jamal, S. B., Naeem, M., Khan, A., & Ghaffar, R. (2013). Phytochemical analysis of medicinal plants occurring in local area of Mardan. *Biochem Anal Biochem*, 2(4), 1-4.
- Wakeel, A., Jan, S. A., Ullah, I., Shinwari, Z. K., & Xu, M. (2019). Solvent polarity mediates phytochemical yield and antioxidant capacity of *Isatis tinctoria*. *PeerJ*, 7, e7857.
- Wang, G. G., & Wangen, S. R. (2011). Does frequent burning affect longleaf pine (*Pinus palustris*) bark thickness?. *Canadian journal of forest research*, 41(7), 1562-1565.
- Whiting, D., Tolan, R., Mecham, B., & Bauer, M. (2003). *Mulching with wood/bark chips, grass clippings, and rock* (Doctoral dissertation, Colorado State University. Libraries).
- Wijaya, T. H., & Ekowati, N. (2021). The Antibacterial Activity of *Acanthus ilicifolius* L. n-Hexane Fraction. *Journal of Science and Technology Research for Pharmacy*, 1(2), 48-56.
- Wynberg, R., & Laird, S. (2007). Bioprospecting: tracking the policy debate. *Environment: Science and Policy for Sustainable Development*, 49(10), 20-32.
- Yang, C. H., Li, R. X., & Chuang, L. Y. (2012). Antioxidant activity of various parts of *Cinnamomum cassia* extracted with different extraction methods. *Molecules*, 17(6), 7294-7304.

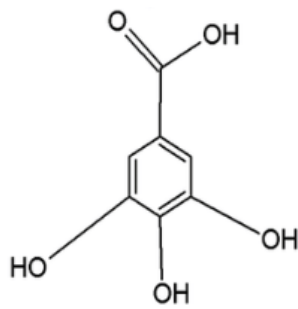
- Yang, W., Chen, X., Li, Y., Guo, S., Wang, Z., & Yu, X. (2020). Advances in pharmacological activities of terpenoids. *Natural Product Communications*, 15(3), 1934578X20903555.
- Ye, Z. W., Zhang, J., Townsend, D. M., & Tew, K. D. (2015). Oxidative stress, redox regulation and diseases of cellular differentiation. *Biochimica et Biophysica Acta (BBA)-General Subjects*, 1850(8), 1607-1621.
- Yeo, Y. L., Chia, Y. Y., Lee, C. H., Sow, H. S., & Yap, W. S. (2014). Effectiveness of maceration periods with different extraction solvents on in-vitro antimicrobial activity from fruit of *Momordica charantia* L. *Journal of Applied Pharmaceutical Science*, 4(10), 16-23.
- Yin, X., Chen, K., Cheng, H., Chen, X., Feng, S., Song, Y., & Liang, L. (2022). Chemical Stability of Ascorbic Acid Integrated into Commercial Products: A Review on Bioactivity and Delivery Technology. *Antioxidants*, 11(1), 153.
- Yusnawan, E. (2013). The effectiveness of polar and non polar fractions of *Ageratum conyzoides* L. to control peanut rust disease and phytochemical screenings of secondary metabolites. *Jurnal Hama dan Penyakit Tumbuhan Tropika*, 13(2), 159-166.
- Zhang, H., Birch, J., Pei, J., Mohamed Ahmed, I. A., Yang, H., Dias, G., ... & Bekhit, A. E. D. (2019). Identification of six phytochemical compounds from *Asparagus officinalis* L. root cultivars from New Zealand and China using UAE-SPE-UPLC-MS/MS: effects of extracts on H₂O₂-induced oxidative stress. *Nutrients*, 11(1), 107.
- Zhang, L., Chen, J., Wang, Y., Wu, D., & Xu, M. (2010). Phenolic extracts from *Acacia mangium* bark and their antioxidant activities. *Molecules*, 15(5), 3567-3577.
- Zhang, Q. W., Lin, L. G., & Ye, W. C. (2018). Techniques for extraction and isolation of natural products: A comprehensive review. *Chinese medicine*, 13(1), 1-26.
- Zoremsiami, J., & Jagetia, G. C. (2018). The Phytochemical and Thin Layer Chromatography Profile of Ethnomedicinal Plant *Helicia Nilagirica* (Bedd). *Int J Pharmacog Chinese Med*, 2(2), 000131.

APPENDICES

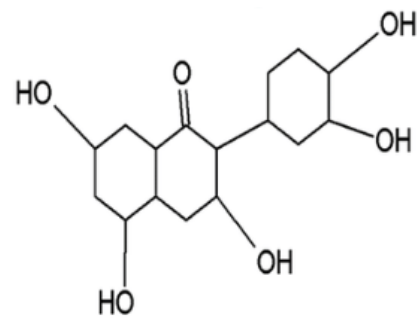
Alkaloids



Appendix 1. Chemical structure of some alkaloids

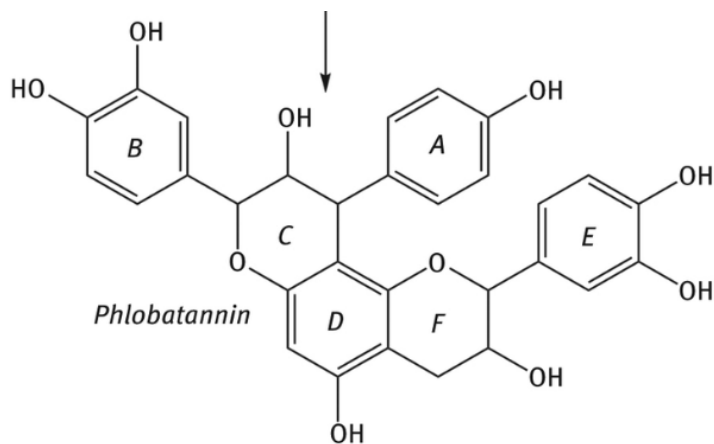


Hydrolysable Tannins

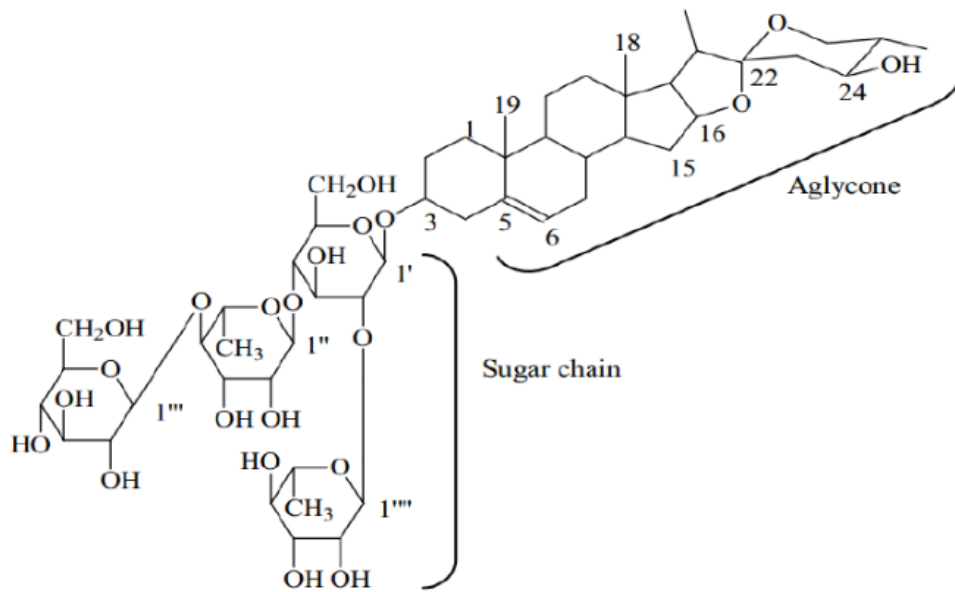


Condensed Tannins

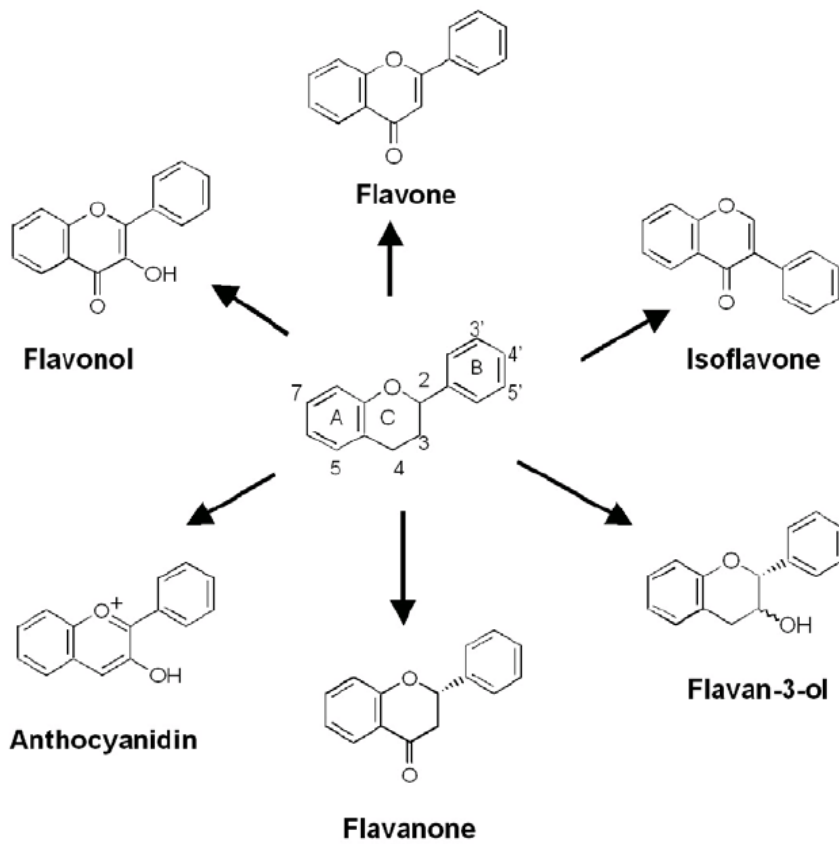
Appendix 2. Chemical structure of hydrolysable tannins (derived from gallic acid) and condensed tannins (resulted from condensation of phenolic compounds)



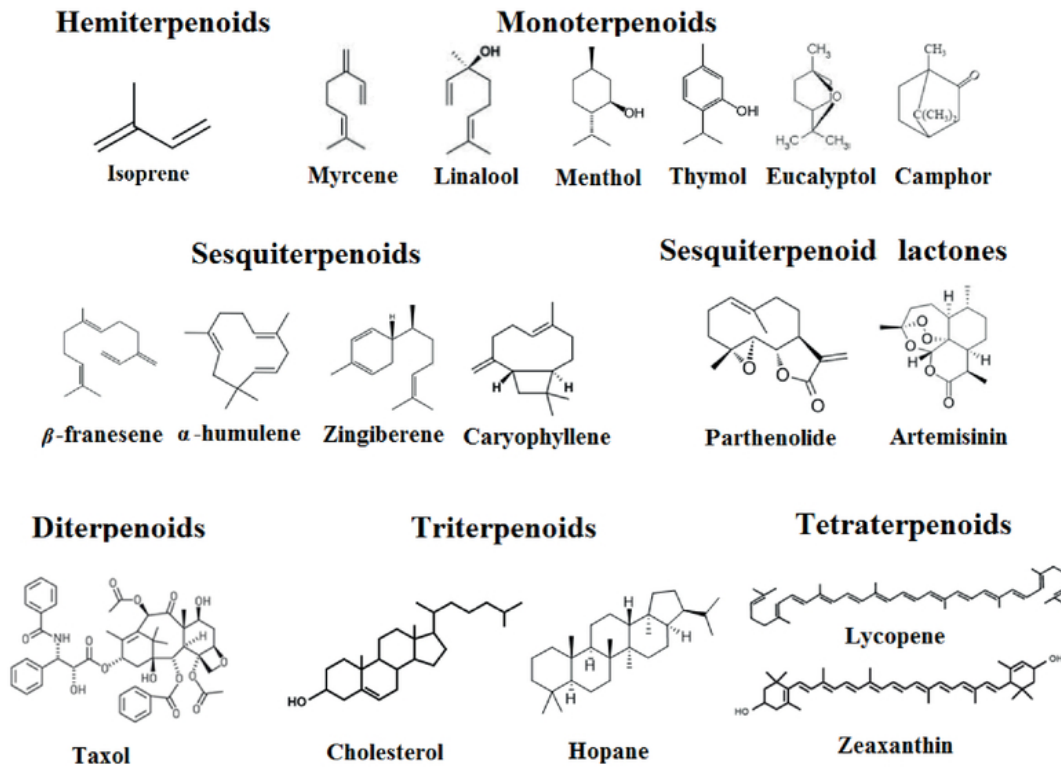
Appendix 3. Chemical structure of phlobatannin



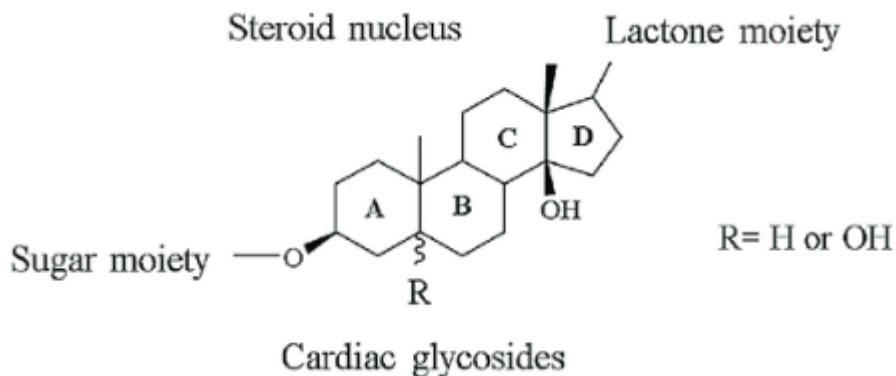
Appendix 4. Chemical structure of saponin



Appendix 5. Basic chemical structure of flavonoids along with chemical structure from its subgroups



Appendix 6. Chemical structure of different classes of terpenoids



Appendix 7. Chemical structure of cardiac glycosides

Concentration (ppm)	Ascorbic Acid					
	First biological replicate			Second biological replicate		
3.90625	0.1478	0.1477	0.151			
7.8125	0.1443	0.1453	0.1438	0.1885	0.1966	0.195
15.625	0.1321	0.1323	0.1354	0.1763	0.1908	0.175
31.25	0.1007	0.1116	0.1115	0.1628	0.1668	0.1675

62.5	0.0691	0.0695	0.0698	0.111	0.1031	0.1108
125	0.0587	0.0574	0.0572	0.0736	0.0721	0.0719
250	0.0494	0.049	0.0494	0.0665	0.0652	0.068
500	0.0525	0.0505	0.0508	0.0617	0.0622	0.061
1,000	0.0479	0.0484	0.0485	0.0544	0.0552	0.0538
Blank	0.1598	0.1597	0.1594	0.1976	0.1977	0.1976

Appendix 8. Raw data for DPPH assay of Ascorbic Acid (First and second biological replicates)

Concentration (ppm)	Methanol					
	First biological replicate			Second biological replicate		
19.53125	0.1917	0.1904	0.1888	0.1736	0.1718	0.1768
39.0625	0.1736	0.167	0.1751	0.166	0.1653	0.1705
78.125	0.1635	0.1543	0.1558	0.1312	0.1513	0.1463
156.25	0.1253	0.1392	0.131	0.1094	0.1116	0.1098
312.5	0.1178	0.1236	0.1118	0.1013	0.1077	0.107
625	0.0878	0.0928	0.091	0.0952	0.1013	0.0951
1250	0.061	0.0649	0.0605	0.0842	0.801	0.0859
2500	0.0535	0.0557	0.0544	0.0679	0.0651	0.0704
Blank	0.194	0.1943	0.1942	0.1768	0.1767	0.1767

Appendix 9. Raw data for DPPH assay of “Kamlowelen” methanol extract (First and second biological replicates)

Concentration (ppm)	Ethyl acetate					
	First biological replicate			Second biological replicate		
156.25	0.1619	0.1664	0.1661	0.1726	0.175	0.173
312.5	0.1645	0.1623	0.1655	0.1679	0.1646	0.1649
625	0.1567	0.1593	0.1563	0.1461	0.1657	0.1746
1,250	0.1527	0.1526	0.1545	0.1574	0.157	0.1584
2,500	0.146	0.1503	0.1454	0.1476	0.1473	0.141

5,000	0.1367	0.1357	0.1389	0.1473	0.1406	0.147
10,000	0.1235	0.1214	0.1207	0.1235	0.1206	0.1344
20,000	0.1019	0.1014	0.1025	0.0921	0.0773	0.0773
Blank	0.1671	0.1669	0.1666	0.1761	0.1760	0.1765

Appendix 10. Raw data for DPPH assay of “Kamlowelen” ethyl acetate extract (First and second biological replicates)

Concentration (ppm)	Chloroform					
	First biological replicate			Second biological replicate		
156.25	0.1564	0.1546	0.1584	0.1351	0.1538	0.1453
312.5	0.1167	0.123	0.1242	0.1432	0.1495	0.139
625	0.1122	0.1126	0.1157	0.1267	0.1245	0.1279
1,250	0.1053	0.1036	0.1039	0.1095	0.0938	0.1167
2,500	0.0845	0.088	0.0878	0.0976	0.0936	0.1005
5,000	0.0786	0.0792	0.0834	0.0894	0.0931	0.0836
10,000	0.0754	0.0752	0.0726	0.0808	0.0808	0.0824
20,000	0.0595	0.063	0.0594	0.0645	0.0637	0.0622
Blank	0.1591	0.1590	0.1589	0.1593	0.1593	0.1592

Appendix 11. Raw data for DPPH assay of “Kamlowelen” chloroform extract (First and second biological replicates)

Concentration (ppm)	Acetone					
	First biological replicate			Second biological replicate		
156.25	0.127	0.1254	0.1265	0.1426	0.1419	0.1423
312.5	0.1257	0.1266	0.1234	0.1325	0.1321	0.1388
625	0.1233	0.1231	0.1242	0.1193	0.1148	0.1127
1,250	0.1138	0.1173	0.1159	0.1112	0.117	0.1153
2,500	0.0934	0.0925	0.0948	0.096	0.0919	0.0981
5,000	0.0804	0.0836	0.0777	0.0881	0.089	0.0895

10,000	0.0786	0.0815	0.0786	0.069	0.0673	0.07
20,000	0.0533	0.0543	0.0533	0.0597	0.0607	0.0642
Blank	0.1271	0.1270	0.1271	0.1448	0.1448	0.1448

Appendix 12. Raw data for DPPH assay of “Kamlowelen” acetone extract (First and second biological replicates)

Concentration (ppm)	Hexane					
	First biological replicate			Second biological replicate		
156.25	0.1789	0.1809	0.1804	0.1758	0.1763	0.1754
312.5	0.1711	0.176	0.1793	0.1722	0.1708	0.1734
625	0.1669	0.1674	0.1625	0.1694	0.1661	0.1757
1,250	0.157	0.1598	0.1539	0.1662	0.1683	0.1657
2,500	0.144	0.1432	0.1485	0.1677	0.1603	0.1605
5,000	0.1089	0.1103	0.107	0.1473	0.1496	0.1494
10,000	0.1088	0.116	0.1056	0.1366	0.1379	0.1375
20,000	0.0624	0.0717	0.0565	0.1243	0.1234	0.1118
Blank	0.1816	0.1815	0.1814	0.1767	0.1766	0.1763

Appendix 13. Raw data for DPPH assay of “Kamlowelen” hexane extract (First and second biological replicates)

Extract	IC50 value
Methanol	496.50 ± 60.29
Ethyl acetate	32777.98 ± 1384.65
Chloroform	5623.23 ± 926.18
Acetone	15218.77 ± 1462.48
Hexane	12132.26 ± 1525.42

Appendix 14. DPPH radical scavenging capabilities of “Kamlowelen” extracted with various solvent in the form of IC50 values

Concentration (ppm)	Methanol					
	First biological replicate			Second biological replicate		
156.25	0.1235	0.1319	0.1269	0.1781	0.1828	0.1848
312.5	0.1232	0.1249	0.1259	0.1703	0.1732	0.1787
625	0.1182	0.1188	0.1247	0.1669	0.1644	0.1658
1,250	0.1025	0.1016	0.107	0.153	0.1543	0.1564
2,500	0.0976	0.0996	0.0951	0.1255	0.1299	0.1264
5,000	0.0683	0.0693	0.0661	0.0982	0.1014	0.1045
10,000	0.0521	0.0563	0.0557	0.0769	0.0803	0.0793
20,000	0.0489	0.0509	0.0527	0.0542	0.0577	0.0578
Blank	0.2148	0.2148	0.2149	0.3245	0.3246	0.3248

Appendix 15. Raw data for DPPH assay of “Kayu Bawang” methanol extract (First and second biological replicates)

Concentration (ppm)	Ethyl acetate					
	First biological replicate			Second biological replicate		
156.25	0.1746	0.1794	0.1807	0.2123	0.2137	0.2135
312.5	0.152	0.1644	0.159	0.1941	0.1948	0.1936
625	0.1493	0.153	0.1519	0.194	0.1877	0.1923
1,250	0.1371	0.1437	0.1332	0.1792	0.1824	0.181
2,500	0.126	0.127	0.127	0.1736	0.1726	0.1777
5,000	0.117	0.1189	0.1128	0.1737	0.1733	0.1711
10,000	0.0695	0.0674	0.0673	0.1459	0.1499	0.1498
20,000	0.0524	0.052	0.0527	0.0965	0.101	0.1011
Blank	0.2193	0.2194	0.2192	0.3183	0.3187	0.3182

Appendix 16. Raw data for DPPH assay of “Kayu Bawang” ethyl acetate extract (First and second biological replicates)

Concentration (ppm)	Chloroform					
	First biological replicate			Second biological replicate		
156.25	0.1497	0.1514	0.151	0.1912	0.1886	0.1945
312.5	0.1361	0.1462	0.147	0.1811	0.1888	0.1877
625	0.1317	0.131	0.1352	0.1739	0.1715	0.1715
1,250	0.1238	0.1285	0.1255	0.1585	0.1605	0.1628
2,500	0.116	0.1178	0.1223	0.145	0.1539	0.1408
5,000	0.0981	0.1005	0.0988	0.1171	0.1398	0.1419
10,000	0.0927	0.0979	0.095	0.1246	0.1292	0.1225
20,000	0.068	0.0795	0.0728	0.1226	0.1246	0.0675
Blank	0.2225	0.2225	0.2228	0.3148	0.3147	0.315

Appendix 17. Raw data for DPPH assay of “Kayu Bawang” chloroform extract (First and second biological replicates)

Concentration (ppm)	Acetone					
	First biological replicate			Second biological replicate		
156.25	0.2205	0.2212	0.2201	0.1869	0.1816	0.1764
312.5	0.177	0.1767	0.1753	0.1934	0.1842	0.1636
625	0.1634	0.1718	0.1709	0.1726	0.174	0.1691
1,250	0.1527	0.162	0.1588	0.1719	0.1682	0.1583
2,500	0.1437	0.1534	0.1544	0.1575	0.1628	0.1616
5,000	0.1369	0.1311	0.1296	0.1158	0.1299	0.1252
10,000	0.0927	0.0933	0.0957	0.1065	0.1041	0.1062
20,000	0.0783	0.0776	0.0773	0.1081	0.105	0.1009
Blank	0.2311	0.2311	0.2311	0.2958	0.2957	0.2955

Appendix 18. Raw data for DPPH assay of “Kayu Bawang” acetone extract (First and second biological replicates)

Concentration (ppm)	Hexane					
	First biological replicate			Second biological replicate		
156.25	0.2219	0.2208	0.2245	0.2111	0.223	0.2199
312.5	0.2164	0.2152	0.2211	0.1974	0.1985	0.1949
625	0.2146	0.2104	0.2156	0.1983	0.1964	0.185
1,250	0.2132	0.2133	0.2143	0.1907	0.1899	0.18
2,500	0.1046	0.2092	0.2031	0.1897	0.188	0.1743
5,000	0.2042	0.203	0.1981	0.1879	0.1877	0.1709
10,000	0.185	0.1856	0.1871	0.183	0.1857	0.1705
20,000	0.12	0.1482	0.1563	0.1767	0.1753	0.1694
Blank	0.2251	0.2252	0.2255	0.3139	0.314	0.3136

Appendix 19. Raw data for DPPH assay of “Kayu Bawang” hexane extract (First and second biological replicates)

Extract	IC50 value
Methanol	1166.08 ± 71.75
Ethyl acetate	4296.23 ± 386.92
Chloroform	3514.16 ± 87.74
Acetone	12614.57 ± 2307.03
Hexane	34141.18 ± 12926.17

Appendix 20. DPPH radical scavenging capabilities of “Kayu Bawang” extracted with various solvent in the form of IC50 values

Concentration (ppm)	Absorbance		Total Phenolic Content (mg GAE / g)	
	(1 st replicate)	(2 nd replicate)	(1 st replicate)	(2 nd replicate)
GA 1.171875	0.069	0.062		
GA 2.34375	0.083	0.127		
GA 4.6875	0.111	0.11		
GA 9.375		0.166		

GA 18.75	0.332	0.293		
GA 37.5	0.535	0.517		
GA 75	0.952	0.924		
GA150	1.634	1.505		
GA 300	3.13	2.884		
KKM 1 625	0.876	0.747	142.49 ± 33.95	115.08 ± 4.51
KKM 2 625	0.91	0.8		
KKM 3 625	1.265	0.77		
KKE 1 20,000	0.953	0.895	4.32 ± 0.31	4.33 ± 0.08
KKE 2 20,000	1.062	0.927		
KKE 3 20,000	0.954	0.912		
KKC 1 1250	1.179	1.114	83.03 ± 4.14	79.54 ± 6.02
KKC 2 1250	1.212	1.007		
KKC 3 1250	1.109	0.976		
KKA 1 5,000	0.861	0.788	14.78 ± 0.33	14.33 ± 1.08
KKA 2 5,000	0.881	0.712		
KKA 3 5,000	0.848	0.809		
KKH 1 20,000	0.427	0.453	1.68 ± 0.19	2.16 ± 0.34
KKH 2 20,000	0.437	0.477		
KKH 3 20,000	0.499	0.573		

Appendix 21. Raw data for Total Phenolic Content analysis of “Kamlowelen” along with Gallic Acid as the standard

Parameter	Value (1 st replicate)	Value (2 nd replicate)
R ²	0.9969	0.9955
Equation	Y = 0.01026*X + 0.08868	Y = 0.009426*X + 0.09437

Appendix 22. R squared and equation from TPC analysis of “Kamlowelen” along with Gallic Acid as the standard

Concentration (ppm)	Absorbance		Total Phenolic Content (mg GAE / g)	
	(1 st replicate)	(2 nd replicate)	(1 st replicate)	(2 nd replicate)
GA 1.171875	0.069	0.062		
GA 2.34375	0.083	0.127		
GA 4.6875	0.111	0.11		
GA 9.375		0.166		
GA 18.75	0.332	0.293		
GA 37.5	0.535	0.517		
GA 75	0.952	0.924		
GA150	1.634	1.505		
GA 300	3.13	2.884		
KBM 1 20,000	3.072	2.845	14.44 ± 0.15	14.97 ± 0.54
KBM 2 20,000	3.016	2.872		
KBM 3 20,000	3.067	3.033		
KBE 1 20,000	0.703	0.32	3 ± 0.56	1.01 ± 0.21
KBE 2 20,000	0.59	0.242		
KBE 3 20,000	0.819	0.293		
KBC 1 20,000	0.229	0.232	0.61 ± 0.12	0.69 ± 0.08
KBC 2 20,000	0.226	0.207		
KBC 3 20,000	0.185	0.32		
KBA 1 20,000	1.741	2.371	8.15 ± 0.16	11.89 ± 0.21
KBA 2 20,000	1.801	2.291		
KBA 3 20,000	1.744	2.343		
KBH 1 20,000	0.149	0.149	0.43 ± 0.18	0.27 ± 0.04
KBH 2 20,000	0.165	0.151		
KBH 3 20,000	0.22	0.136		

Appendix 23. Raw data for TPC analysis of “Kayu Bawang” along with Gallic Acid as the standard

Parameter	Value (1 st replicate)	Value (2 nd replicate)
R ²	0.9969	0.9955
Equation	Y = 0.01026*X + 0.08868	Y = 0.009426*X + 0.09437

Appendix 24. R squared and equation from TPC analysis of “Kayu Bawang” along with Gallic Acid as the standard

Feedback Studio - Google Chrome
 ev.turnitin.com/app/carta/en_us/?lang=en_us&u=1087771463&student_user=1&s=1&o=1849675106

feedback studio Gabriela Lysette | Final research report

Match Overview

6%

- 1 bioresources.cnr.ncsu... <1% >
Internet Source
- 2 Submitted to University... <1% >
Student Paper
- 3 Kevser Karaman, "Volat... <1% >
Publication
- 4 Sónia A.O. Santos, Paul... <1% >
Publication
- 5 www.mdpi.com <1% >
Internet Source
- 6 umpir.ump.edu.my <1% >
Internet Source
- 7 Zoltan Pásztor, Ildikó ... <1% >
Publication

ABSTRACT

“Kamlowelen” and “Kayu Bawang” are native papuan trees that have never been investigated before. Both tree barks are commonly used as a seasoning, tea, and medicine. In the present study, total phenolic content (TPC), DPPH assay, and phytochemical screening of tree barks extracted using five different solvents (Methanol, acetone, ethyl acetate, chloroform, and hexane) were analyzed. Among five solvents, methanol extract exhibit the lowest IC50 values (496.5±60.29 µg extract/ml at “Kamlowelen” and 1166.08±71.75 µg extract/ml for “Kayu Bawang”), while the highest IC50 values was found in hexane extract (50763.61±12299.76 µg extract/ml for “Kamlowelen” and 42694.59±11535.01 µg extract/ml for “Kayu Bawang”). In addition, the highest total phenolic activity was also observed in methanol extract for both tree barks (142.49±33.96 mg GAE/g extracts for “Kamlowelen” and 14.97±0.54 mg GAE/g extracts for “Kayu Bawang”). As for the phytochemical screening, methanol and acetone extract showed the best result as they were able to extract the most phytochemical compounds classes compared to other solvent. The result of this study showed that the choice of solvent for extraction can

Page: 1 of 28 Word Count: 8935 Text-Only Report High Resolution On 9:45 PM 6/3/2022

Appendix 25. Plagiarism checker of final thesis report