

References

- Aljabali, A. A., El-Tanani, M., & Tambuwala, M. M. (2024). Principles of CRISPR-Cas9 technology: Advancements in genome editing and emerging trends in drug delivery. *Journal of Drug Delivery Science and Technology*, 92, 105338. <https://doi.org/10.1016/j.jddst.2024.105338>
- Ansori, A. N., Antonius, Y., Susilo, R. J., Hayaza, S., Kharisma, V. D., Parikesit, A. A., Zainul, R., Jakhmola, V., Saklani, T., Rebezov, M., Ullah, M. E., Maksimiuk, N., Derkho, M., & Burkov, P. (2023). Application of CRISPR-Cas9 genome editing technology in various fields: A review. *Narra J*, 3(2), e184. <https://doi.org/10.52225/narra.v3i2.184>
- Bhattacharya, S., & Satpati, P. (2022). Insights into the Mechanism of CRISPR/Cas9-Based Genome Editing from Molecular Dynamics Simulations. *ACS Omega*, 8(2), 1817–1837. <https://doi.org/10.1021/acsomega.2c05583>
- Chahal, G., Helbig, K., Parton, R., & Monson, E. (2026). The Biology of Endosomal Escape: Strategies for Enhanced Delivery of Therapeutics. *ACS Nano*, 20(2), 1789–1813. <https://doi.org/10.1021/acsnano.5c18112>
- Chen Y. (2011). Calcium Phosphate Transfection of Eukaryotic Cells. *Bio-protocol*, 1(7). <https://static.igem.org/mediawiki/2018/c/c8/T--NYMU-Taipei--protocol-transfection.pdf>
- Chen, W., McKenna, A., Schreiber, J., Haeussler, M., Yin, Y., Agarwal, V., Noble, W. S., & Shendure, J. (2019). Massively parallel profiling and predictive modeling of the outcomes of CRISPR/Cas9-mediated double-strand break repair. *Nucleic Acids Research*, 47(15), 7989–8003. <https://doi.org/10.1093/nar/gkz487>
- Chong, Z. X., Yeap, S. K., & Ho, W. Y. (2021). Transfection types, methods and strategies: a technical review. *PeerJ*, 9, e11165. <https://doi.org/10.7717/peerj.11165>
- Concordet, J., & Haeussler, M. (2018). CRISPOR: intuitive guide selection for CRISPR/Cas9 genome editing experiments and screens. *Nucleic Acids Research*, 46(W1), W242–W245. <https://doi.org/10.1093/nar/gky354>
- Conese, M. (2009). Polyethylenimine-mediated gene delivery to the lung and therapeutic applications. *Drug Design Development and Therapy*, 2, 163. <https://doi.org/10.2147/dddt.s2708>
- Curtsinger, H. D., Martínez-Absalón, S., Liu, Y., & Lopatkin, A. J. (2024). The metabolic burden associated with plasmid acquisition: An assessment of the unrecognized benefits to host cells. *BioEssays*, 47(2), e2400164. <https://doi.org/10.1002/bies.202400164>
- Dabbaghi, M., Hashemi, K., Oskuee, R. K., & Afkhami-Goli, A. (2022). Reverse relation between cytotoxicity and Polyethylenimine/DNA ratio, the effect of using HEPES-buffered saline (HBS)

- medium in gene delivery. *Toxicology in Vitro*, 83, 105414. <https://doi.org/10.1016/j.tiv.2022.105414>
- Dersch, S., & Graumann, P. L. (2024). Adaptation of *Bacillus subtilis* MreB Filaments to Osmotic Stress Depends on Influx of Potassium Ions. *Microorganisms*, 12(7), 1309. <https://doi.org/10.3390/microorganisms12071309>
- Ding, S., Liu, J., Han, X., & Tang, M. (2023). CRISPR/CAS9-Mediated Genome editing in cancer therapy. *International Journal of Molecular Sciences*, 24(22), 16325. <https://doi.org/10.3390/ijms242216325>
- Ding, M., Ma, C., Lin, Y., Fang, H., Xu, Y., Wang, S., Chen, Y., Zhou, J., Gao, H., Shan, Y., Yang, L., Sun, H., Tang, Y., Wu, X., Zhu, L., Zheng, L., Assaraf, Y. G., Zhou, B. S., Gu, S., & Li, H. (2025). Therapeutic targeting de novo purine biosynthesis driven by β -catenin-dependent PPAT upregulation in hepatoblastoma. *Cell Death and Disease*, 16(1), 179. <https://doi.org/10.1038/s41419-025-07502-6>
- Doench, J. G., Hartenian, E., Graham, D. B., Tothova, Z., Hegde, M., Smith, I., Sullender, M., Ebert, B. L., Xavier, R. J., & Root, D. E. (2014). Rational design of highly active sgRNAs for CRISPR-Cas9-mediated gene inactivation. *Nature Biotechnology*, 32(12), 1262–1267. <https://doi.org/10.1038/nbt.3026>
- Doench, J. G., Fusi, N., Sullender, M., Hegde, M., Vaimberg, E. W., Donovan, K. F., Smith, I., Tothova, Z., Wilen, C., Orchard, R., Virgin, H. W., Listgarten, J., & Root, D. E. (2016). Optimized sgRNA design to maximize activity and minimize off-target effects of CRISPR-Cas9. *Nature Biotechnology*, 34(2), 184–191. <https://doi.org/10.1038/nbt.3437>
- Erkmann, J. (2004). Nuclear export of mRNA: from the site of transcription to the cytoplasm. *Experimental Cell Research*, 296(1), 12–20. <https://doi.org/10.1016/j.yexcr.2004.03.015>
- Falah, G., Sharvit, L., & Atzmon, G. (2024). CRISPR-Cas9 mediated d3GHR knockout in HEK293 cells: Revealing the longevity associated isoform stress resilience. *Experimental Gerontology*, 196, 112586. <https://doi.org/10.1016/j.exger.2024.112586>
- Fus-Kujawa, A., Prus, P., Bajdak-Rusinek, K., Teper, P., Gawron, K., Kowalczyk, A., & Sieron, A. L. (2021). An Overview of Methods and Tools for Transfection of Eukaryotic Cells in vitro. *Frontiers in Bioengineering and Biotechnology*, 9, 701031. <https://doi.org/10.3389/fbioe.2021.701031>
- Geneaid Biotech Ltd. (n.d.). Presto™ Mini Plasmid Kit. <https://www.geneaid.com/data/files/1608174169928361233.pdf>
- Gostimskaya, I. (2022). CRISPR–CAS9: A history of its discovery and ethical considerations of its use in genome editing. *Biochemistry (Moscow)*, 87(8), 777–788. <https://doi.org/10.1134/s0006297922080090>

- Halid, N. A. M. A., Yern, K. K., Dana, A., Zain, N. A. M. (2021). Determination of In Vitro Synergy of Ampicillin and Chloramphenicol against Multidrug Resistant *Bacillus cereus* Species. *International Journal of Life Sciences and Biotechnology*, 5(1), 42–55. <https://doi.org/10.38001/ijlsb.970670>
- Hasan, M. M., Ragnarsson, L., Cardoso, F. C., & Lewis, R. J. (2021). Transfection methods for high-throughput cellular assays of voltage-gated calcium and sodium channels involved in pain. *Plos one*, 16(3), e0243645.
- Hayashi, H., Kubo, Y., Izumida, M., & Matsuyama, T. (2020). Efficient viral delivery of Cas9 into human safe harbor. *Scientific Reports*, 10(1). <https://doi.org/10.1038/s41598-020-78450-8>
- Hermantara, R., Richmond, L., Taqi, A. F., Chilaka, S., Jeantet, V., Guerrini, I., West, K., & West, A. (2024). Improving CRISPR–Cas9 directed faithful transgene integration outcomes by reducing unwanted random DNA integration. *Journal of Biomedical Science*, 31(1), 32. <https://doi.org/10.1186/s12929-024-01020-x>
- Holban, A.-M., & Grumezescu, A. (2019). *Materials for biomedical engineering: Hydrogels and polymer-based scaffolds*. Elsevier.
- Horizon Discovery. (2025). *T7EI calculator*. Revvity Discovery Limited. Retrieved July 11, 2026, from <https://horizondiscovery.com/en/ordering-and-calculation-tools/t7ei-calculator>
- Hosseini-Kharat, M., Bremmell, K. E., Grubor-Bauk, B., & Prestidge, C. A. (2025). Enhancing non-viral DNA delivery systems: Recent advances in improving efficiency and target specificity. *Journal of Controlled Release*, 378, 170–194. <https://doi.org/10.1016/j.jconrel.2024.12.002>
- Hsu, P. D., Scott, D. A., Weinstein, J. A., Ran, F. A., Konermann, S., Agarwala, V., Li, Y., Fine, E. J., Wu, X., Shalem, O., Cradick, T. J., Marraffini, L. A., Bao, G., & Zhang, F. (2013). DNA targeting specificity of RNA-guided Cas9 nucleases. *Nature Biotechnology*, 31(9), 827–832. <https://doi.org/10.1038/nbt.2647>
- Invitrogen Corporation. (2007). PureLink™ Genomic DNA Mini Kit. https://documents.thermofisher.com/TFS-Assets/LSG/manuals/purelink_genomic_mini_man.pdf
- Jahanafrooz, Z., Bakhshandeh, B., & Shirzadi, E. (2022). Comparative evaluation of lipofectamine and dendrimer for transfection of short RNA into human T47D and MCF-10A cell lines. *Advanced Pharmaceutical Bulletin*, 13(2), 385–392. <https://doi.org/10.34172/apb.2023.022>
- Jaiswal, R., Braud, B., Hernandez-Ramirez, K. C., Santosh, V., Washington, A., & Escalante, C. R. (2025). Cryo-EM structure of AAV2 Rep68 bound to integration site AAVS1: insights into the mechanism of DNA melting. *Nucleic Acids Research*, 53(3). <https://doi.org/10.1093/nar/gkaf033>

- Javaid, D., Ganie, S. Y., Hajam, Y. A., & Reshi, M. S. (2022). CRISPR/Cas9 system: A reliable and facile genome editing tool in modern biology. *Molecular Biology Reports*. <https://doi.org/10.1007/s11033-022-07880-6>
- Jena, R., Vishwas, S., Kumar, R., Kaur, J., Khursheed, R., Gulati, M., Singh, T. G., Vanathi, B. M., Alam, A., Kumar, B., Chaitanya, M. V. N. L., Gupta, S., Negi, P., Pandey, N. K., Bhatt, S., Gupta, G., Chellappan, D. K., Oliver, B. G., Dua, K., & Singh, S. K. (2022). Treatment strategies for HIV infection with emphasis on role of CRISPR/Cas9 gene: Success so far and road ahead. *European Journal of Pharmacology*, 931, 175173. <https://doi.org/10.1016/j.ejphar.2022.175173>
- Jin, Y., Zheng, H., Ibanez, A. C. S., Patil, P. D., Lv, S., Luo, M., Duncan, T. M., & Luk, Y. (2019). Cell-Wall-Targeting antibiotics cause Lag-Phase bacteria to form Surface-Mediated filaments promoting the formation of biofilms and aggregates. *ChemBioChem*, 21(6), 825–835. <https://doi.org/10.1002/cbic.201900508>
- Li, D., Liu, J., Sun, L., Zhang, J., & Hou, J. (2024). Developing polycistronic expression tool in *Yarrowia lipolytica*. *Synthetic and Systems Biotechnology*, 10(1), 127–132. <https://doi.org/10.1016/j.synbio.2024.09.010>
- Liu, J., Chang, W., Pan, L., Liu, X., Su, L., Zhang, W., Li, Q., & Zheng, Y. (2018). An Improved Method of Preparing High Efficiency Transformation *Escherichia coli* with Both Plasmids and Larger DNA Fragments. *Indian Journal of Microbiology*, 58(4), 448–456. <https://doi.org/10.1007/s12088-018-0743-z>
- Liao, H., Wu, J., VanDusen, N. J., Li, Y., & Zheng, Y. (2024). CRISPR-Cas9-mediated homology-directed repair for precise gene editing. *Molecular Therapy — Nucleic Acids*, 35(4), 102344. <https://doi.org/10.1016/j.omtn.2024.102344>
- Lu, J., Wu, T., Zhang, B., Liu, S., Song, W., Qiao, J., & Ruan, H. (2021). Types of nuclear localization signals and mechanisms of protein import into the nucleus. *Cell Communication and Signaling*, 19(1), 60. <https://doi.org/10.1186/s12964-021-00741-y>
- Kaur, A., Assogba, V. A., Assogba, J. M. G., & Assogba, B. D. (2025). Optimizing transfection efficiency in primary human splenic fibroblast cells. *bioRxiv* (Cold Spring Harbor Laboratory). <https://doi.org/10.1101/2025.01.19.633792>
- Kocher, T., Peking, P., Klausegger, A., Murauer, E. M., Hofbauer, J. P., Wally, V., Lettner, T., Hainzl, S., Ablinger, M., Bauer, J. W., Reichelt, J., & Koller, U. (2017). Cut and Paste: Efficient Homology-Directed Repair of a Dominant Negative KRT14 Mutation via CRISPR/Cas9 Nickases. *Molecular Therapy*, 25(11), 2585–2598. <https://doi.org/10.1016/j.ymthe.2017.08.015>

- Kurniati, D. E., Riani, C., Hardiyanti, R., & Rachmawati, D. (2025). Optimization of PCR primers for detection of Extended-Spectrum Beta-Lactamase targeting CTX-M and TEM genes. *3BIO Journal of Biological Science Technology and Management*, 7(1), 326–333. <https://doi.org/10.5614/3bio.2025.7.1.5>
- Luft, C., & Ketteler, R. (2015). Electroporation knows no boundaries: the use of electrostimulation for siRNA delivery in cells and tissues. *SLAS DISCOVERY*, 20(8), 932–942. <https://doi.org/10.1177/1087057115579638>
- Madeira, C., Ribeiro, S. C., Turk, M. Z., & Cabral, J. M. S. (2011). Optimization of gene delivery to HEK293T cells by microporation using a central composite design methodology. *Biotechnology Letters*, 32(10), 1393–1399. <https://doi.org/10.1007/s10529-010-0327-4>
- Matlock, B. (n.d.). *Assesment of Nucleic Acid Purity*. ThermoFisher. <https://documents.thermofisher.com/TFS-Assets/CAD/Product-Bulletins/TN52646-E-0215M-NucleicAcid.pdf>
- Mengstie, M. A., & Wondimu, B. Z. (2021). Mechanism and applications of CRISPR/CAS-9-Mediated Genome Editing. *Biologics*, 15, 353–361. <https://doi.org/10.2147/btt.s326422>
- Mengstie, M. A., Azezew, M. T., Dejenie, T. A., Teshome, A. A., Admasu, F. T., Teklemariam, A. B., Mulu, A. T., Agidew, M. M., Adugna, D. G., Geremew, H., & Abebe, E. C. (2024). Recent advancements in reducing the off-target effect of CRISPR-Cas9 genome editing. *Biologics: Targets and Therapy*, 18(1), 21–28. <https://doi.org/10.2147/BTT.S429411>
- Moreno-Mateos, M. A., Vejnár, C. E., Beaudoin, J., Fernandez, J. P., Mis, E. K., Khokha, M. K., & Giraldez, A. J. (2015). CRISPRscan: designing highly efficient sgRNAs for CRISPR-Cas9 targeting in vivo. *Nature Methods*, 12(10), 982–988. <https://doi.org/10.1038/nmeth.3543>
- Mou, J., Shi, H., Chen, Q., Dong, Y., Wang, J., Yang, M., & Zhang, Z. (2026). A short communication: Genome editing and mutation detection of triploid tree induced by CRISPR/Cas9. *Industrial Crops and Products*, 246, 123389. <https://doi.org/10.1016/j.indcrop.2026.123389>
- Ni, J., Gong, J., Ran, Y., Bai, R., Huang, P., Zheng, Z., Zhou, M., Lan, F., Gu, W., & Liu, X. (2025). Optimization of gene knockout approaches and sgRNA selection in hPSCs with inducible Cas9 expression. *Scientific Reports*, 15(1), 40461. <https://doi.org/10.1038/s41598-025-24184-4>
- Oceguera-Yanez, F., Kim, S., Matsumoto, T., Tan, G. W., Xiang, L., Hatani, T., Kondo, T., Ikeya, M., Yoshida, Y., Inoue, H., & Woltjen, K. (2016). Engineering the AAVS1 locus for consistent and scalable transgene expression in human iPSCs and their differentiated derivatives. *Methods*, 101, 43–55. <https://doi.org/10.1016/j.ymeth.2015.12.012>
- QIAGEN. (2008). QIAquick PCR Purification Kit Protocol. https://static.igem.org/mediawiki/2012/a/a3/QIAquick_PCR-purification.pdf

- QIAGEN. (2025). *QIAamp DNA Mini and Blood Mini handbook* (HB-0329-007).
<https://www.qiagen.com/en-US/resources/download/KitHandbook/hb-0329-007-hb-qiaamp-dna-mini-blood-mini-0725-ww>
- Queen's Center for Biomedical Research. (2023). *Protocol for HEK293 Polyethylenimine transfection*.
<https://share.google/Q5HHkBNK99yaRSzM7>
- Rana, N., Panda, A. K., Pathak, N., Gupta, T., & Thakur, S. D. (2020). Bacillus cereus: public health burden associated with ready-to-eat foods in Himachal Pradesh, India. *Journal of Food Science and Technology*, 57(6), 2293–2302. <https://doi.org/10.1007/s13197-020-04267-y>
- Rohatgi, N., Fortin, J., Lau, T., Ying, Y., Zhang, Y., Lee, B. L., Costa, M. R., & Reja, R. (2024). Seed sequences mediate off-target activity in the CRISPR-interference system. *Cell Genomics*, 4(11), 100693. <https://doi.org/10.1016/j.xgen.2024.100693>
- Richards, C., Sarkar, S., Kandell, J., Snyder, R., & Lakshmipathy, U. (2023). Assessing the suitability of cell counting methods during different stages of a cell processing workflow using an ISO 20391-2 guided study design and analysis. *Frontiers in Bioengineering and Biotechnology*, 11, 1223227. <https://doi.org/10.3389/fbioe.2023.1223227>
- Ryu, Y. C., Bao, G., & Hwang, B. H. (2025). Peptide-assisted lipofection enables efficient non-viral delivery of large CRISPR/Cas9 constructs for genome editing applications. *Biomedicine & Pharmacotherapy*, 193, 118838. <https://doi.org/10.1016/j.biopha.2025.118838>
- Safarikova, M., Kubena, A. A., Frankova, V., Zima, T., & Kalousova, M. (2021). The effects of different storage conditions and repeated Freeze/THAW cycles on the concentration, purity and integrity of genomic DNA. *Folia Biologica*, 67(1), 10–15. <https://doi.org/10.14712/fb2021067010010>
- Santos, D. P., Kiskinis, E., Eggan, K., & Merkle, F. T. (2016). Comprehensive protocols for CRISPR/CAS9-based gene editing in human pluripotent stem cells. *Current Protocols in Stem Cell Biology*, 38(1), 5B.6.1-5B.6.60. <https://doi.org/10.1002/cpsc.15>
- Scarfe, J., Kosmützky, D., & Nisbet, R. E. R. (2025). A game of tag: A review of protein tags for the successful detection, purification and fluorescence labelling of proteins expressed in microalgae. *The Plant Journal*, 122(6), e70272. <https://doi.org/10.1111/tpj.70272>
- Setia, A., Patil, D., Randhave, N. V., Vaishali, Verma, N., Rani, K., Kale, A., Wagh, B., Kumar, V., Malik, A. K., Pappu, J. S. M., Deshmukh, K., & Muthu, M. S. (2026). CRISPR/Cas9 genome editing-based preclinical theranostics, biomarkers and drug delivery systems for cancer applications. *International Journal of Biological Macromolecules*, 345, 150582. <https://doi.org/10.1016/j.ijbiomac.2026.150582>

- Shi, B., Xue, M., Wang, Y., Wang, Y., Li, D., Zhao, X., & Li, X. (2018). An improved method for increasing the efficiency of gene transfection and transduction. *International journal of physiology, pathophysiology and pharmacology*, 10(2), 95.
- Shumega, A. R., Pavlov, Y. I., Chirinskaite, A. V., Rubel, A. A., Inge-Vechtomov, S. G., & Stepchenkova, E. I. (2024). CRISPR/CAS9 as a mutagenic factor. *International Journal of Molecular Sciences*, 25(2), 823. <https://doi.org/10.3390/ijms25020823>
- Smesnik, G., Virgolini, N., Toth, M., Dürauer, A., & Borth, N. (2025). Comparative Analysis of HEK293 genomic variability. *Biotechnology and Bioengineering*, 123(2), 436–448. <https://doi.org/10.1002/bit.70105>
- Strzyz, P. (2020). CRISPR–Cas9 wins Nobel. *Nature Reviews Molecular Cell Biology*, 21(12), 714. <https://doi.org/10.1038/s41580-020-00307-9>
- Tabata, S., Umemura, S., Narita, M., Udagawa, H., Ishikawa, T., Tsuboi, M., Goto, K., Ishii, G., Tsuchihara, K., Ochiai, A., Kobayashi, S. S., Soga, T., & Makinoshima, H. (2023). Metabolic hallmarks for purine nucleotide biosynthesis in small cell lung carcinoma. *Molecular Cancer Research*, 22(1), 82–93. <https://doi.org/10.1158/1541-7786.mcr-23-0386>
- Tan, E., Chin, C. S. H., Lim, Z. F. S., & Ng, S. K. (2021). HEK293 cell line as a platform to produce recombinant proteins and viral vectors. *Frontiers in Bioengineering and Biotechnology*, 9, 796991. <https://doi.org/10.3389/fbioe.2021.796991>
- Thapa, B., & Narain, R. (2016). Mechanism, current challenges and new approaches for non viral gene delivery. *Polymers and Nanomaterials for Gene Therapy*, 1–27. <https://doi.org/10.1016/b978-0-08-100520-0.00001-1>
- Tsai, S. Q., & Joung, J. K. (2016). Defining and improving the genome-wide specificities of CRISPR–Cas9 nucleases. *Nature Reviews Genetics*, 17(5), 300–312. <https://doi.org/10.1038/nrg.2016.28>
- Vilain, S., Luo, Y., Hildreth, M. B., & Brožek, V. S. (2006). Analysis of the Life Cycle of the Soil Saprophyte *Bacillus cereus* in Liquid Soil Extract and in Soil. *Applied and Environmental Microbiology*, 72(7), 4970–4977. <https://doi.org/10.1128/aem.03076-05>
- Vona, M., Bodenmann, I., Perrenoud, M., Buchs, R., Yasli, P. K., Nanni, L., Daveau, R., Félix, A., Stolte, J., Hov, A., Chollet, B., Schuepbach, T., Ley, D., Guzman, E., Fisch, I., & Mermod, N. (2026). Generation and characterization of a HEK293 cell line optimized for recombinant adeno-associated virus production. *Biotechnology Reports*, 49, e00948. <https://doi.org/10.1016/j.btre.2026.e00948>
- Wang, T., Wei, J. J., Sabatini, D. M., & Lander, E. S. (2013). Genetic screens in human cells using the CRISPR-CAS9 system. *Science*, 343(6166), 80–84. <https://doi.org/10.1126/science.1246981>

- Yang, F., Fang, H., Wang, D., Chen, Y., Zhai, Y., Zhou, B. S., & Li, H. (2017). HPRT1 activity loss is associated with resistance to thiopurine in ALL. *Oncotarget*, 9(2), 2268–2278. <https://doi.org/10.18632/oncotarget.23405>
- Yao, B., Yang, Q., Gonçalves, M. a. F. V., Schiffelers, R., Sluijter, J. P. G., & Lei, Z. (2025). Comparative Analysis of Methods for Assessing On-Target Gene Editing Efficiencies. *Methods and Protocols*, 8(2), 23. <https://doi.org/10.3390/mps8020023>
- Zapata, A. M. G., Pico, A. C., Gusmão, L., Amorim, A., & Sanabria, F. R. (2016). A new mutation at exon 2 of hprt1 locus causing lesch-nyhan syndrome. *Innovaciencia Facultad De Ciencias Exactas Físicas Y Naturales*, 3(1), 18–21. <https://doi.org/10.15649/2346075x.362>
- Zhang, R., He, Z., Shi, Y., Sun, X., Chen, X., Wang, G., Zhang, Y., Gao, P., Wu, Y., Lu, S., Duan, J., Sun, S., Yang, N., Fan, W., Zhao, K., Yang, B., Xia, Y., Zhang, Y., Zhang, Y., & Yin, H. (2024). Amplification editing enables efficient and precise duplication of DNA from short sequence to megabase and chromosomal scale. *Cell*, 187(15), 3936-3952.e19. <https://doi.org/10.1016/j.cell.2024.05.056>
- Zhou, J., Liu, K., Bauer, C., Bendner, G., Dietrich, H., Slivka, J. P., Wink, M., Wong, M. B. F., Chan, M. K. S., & Skutella, T. (2023). Modulation of Cellular Senescence in HEK293 and HepG2 Cells by Ultrafiltrates UPLa and ULu Is Partly Mediated by Modulation of Mitochondrial Homeostasis under Oxidative Stress. *International Journal of Molecular Sciences*, 24(7), 6748. <https://doi.org/10.3390/ijms24076748>