

References

- Abdalla, M., Khan, J., Aldosari, S., El-Arabey, A., Alshehri, B., Rakshit, G., Eltayeb, W., & Rudrapal, M. (2023). Dual synergistic inhibition of COX and LOX by potential chemicals from Indian daily spices investigated through detailed computational studies. *Scientific Reports*, 13. <https://doi.org/10.1038/s41598-023-35161-0>.
- Abdurakhmonov, I. Y. (2022). Introductory chapter: Model plants for discovering the key biological processes in plant. *Model Organisms in Plant Genetics*, 3. <https://doi.org/10.5772/intechopen.103759>.
- Adebomojo, A. A., & AbdulRahaman, A. A. (2020). Surface sterilization of *Ocimum* seeds and tissues with biosynthesized nanosilver and its effects on callus induction. *IOP Conference Series: Materials Science and Engineering*, 805(1), 012024. IOP Publishing. <https://doi.org/10.1088/1757-899X/805/1/012024>.
- Afriani, R., Mambang, D. E. P., Dalimunthe, G. I., & Nasution, H. M. (2022). Uji aktivitas analgesik ekstrak daun timun tikus (*Coccinia grandis* (L). Voigt) terhadap mencit jantan (*Mus musculus*). *Journal of Health and Medical Science*, 169-176.
- Ahmad, M. F., Ahmad, F. A., Alsayegh, A. A., Zeyauallah, M., AlShahrani, A. M., Muzammil, K., Saati, A. A., Wahab, S., Elbendary, E. Y., Kambal, N., Abdelrahman, M. H., & Hussain, S. (2024). Pesticides impacts on human health and the environment with their mechanisms of action and possible countermeasures. *Heliyon*, 10(7), e29128. <https://doi.org/10.1016/j.heliyon.2024.e29128>.
- Ahmed, N., Perveen, F. F., Akter, M., Mamun, A. A., & Islam, M. N. (2023a). Harnessing the therapeutic potential of *Coccinia grandis* phytochemicals in diabetes: A computational, DFT calculation and MMGBSA perspective on aldose reductase inhibition. *Informatics in Medicine Unlocked*, 46, 101477. <https://doi.org/10.1016/j.imu.2024.101477>.
- Ahmed, N., Zhang, B., Chachar, Z., Li, J., Xiao, G., Wang, Q., Hayat, F., Deng, L., Narejo, M., Bozdar, B., & Tu, P. (2023b). Micronutrients and their effects on Horticultural crop quality, productivity and sustainability. *Scientia Horticulturae*, 323, 112512. <https://doi.org/10.1016/j.scienta.2023.112512>.
- Ahmed, R. M., Widdatallah, M. O., Alrasheid, A. A., Widadallah, H. A., & Alkhawad, A. O. (2024). Evaluation of the phytochemical composition, antioxidant properties, and in vivo antihyperglycemic effect of *Coccinia grandis* leaf extract in mice. *Universal Journal of Pharmaceutical Research*, 9(2). <https://doi.org/10.22270/ujpr.v9i2.1091>.
- Akbar, M., Raza, A., Khalil, T., Yasin, N. A., Nazir, Y., & Ahmad, A. (2023). Isolation of herbicidal compounds, quercetin and β -caryophyllene, from *Digera muricata*. *Arabian Journal of Chemistry*, 16(5), 104653. <https://doi.org/10.1016/j.arabjc.2023.104653>.
- Akter, K. M., Sajib, N. H., Kang, D. M., Ahn, M. J., & Uddin, S. B. (2021). Ethnomedicinal study of plants in Begumganj, Noakhali, Bangladesh. *Natural Product Sciences*, 27(4), 217-227. <https://doi.org/10.20307/nps.2021.27.4.217>.
- Alnasser, M., Al-Qahtani, A., Albrahim, T., Alkahtani, S., Al-Anazi, M., & Alkahtani, M. (2020). Potential anti-inflammatory and anti-apoptotic effect of *Coccinia grandis* plant extract in LPS stimulated-THP-1 cells. *Environmental Science and Pollution Research*, 27, 21892 - 21904. <https://doi.org/10.1007/s11356-020-08445-5>.
- Anwar, T., Qureshi, H., Mahnashi, M. H., Kabir, F., Parveen, N., Ahmed, D., Afzal, U., Batool, S., Awais, M., Ahmed Alyami, S., & Ahmed Alhaider, H. (2021a). Bioherbicidal ability and weed management

- of allelopathic methyl esters from *Lantana camara*. *Saudi Journal of Biological Sciences*, 28(8), 4365-4374. <https://doi.org/10.1016/j.sjbs.2021.04.026>.
- Anwar, S., Naseem, S., Karimi, S., Asi, M. R., Akrem, A., & Ali, Z. (2021b). Bioherbicidal activity and metabolic profiling of potent allelopathic plant fractions against major weeds of wheat—way forward to lower the risk of synthetic herbicides. *Frontiers in Plant Science*, 12, 632390. <https://doi.org/10.3389/fpls.2021.632390>.
- Argüelles, L., & March, H. (2022). Weeds in action: Vegetal political ecology of unwanted plants. *Progress in Human Geography*. https://doi.org/10.1177_03091325211054966.
- Arun, M. N., Kumar, R. M., Sreedevi, B., Padmavathi, G., Revathi, P., Pathak, N., ... & Venkatanna, B. (2022). The rising threat of invasive alien plant species in agriculture. *Resource Management in Agroecosystems*. IntechOpen. <http://dx.doi.org/10.5772/intechopen.106742>.
- Aziza, H., Malaysia, E., Wati Lestari, A., & Ngapiyatun, S. (2024). The effectiveness of herbicide use in weed control in oil palm plantation plate areas. *Buletin Loupe*, 20(02), 122–128. <https://doi.org/10.51967/buletinloupe.v20i02.3170>.
- Balderas-Renterías, I., Arredondo-Espinoza, E., Sánchez-Carranza, J., Ramírez-Estrada, K., Camacho-Corona, M., Carrizales-Castillo, J., Romo-Mancillas, A., Alcantar-Rosales, V., Stefani, T., & González-Maya, L. (2021). Cytotoxic fractions from *Hechtia glomerata* extracts and p-coumaric acid as MAPK inhibitors. *Molecules*, 26. <https://doi.org/10.3390/molecules26041096>.
- Baldrige, D., Wangler, M. F., Bowman, A. N., Yamamoto, S., Schedl, T., Pak, S. C., ... & Westerfield, M. (2021). Model organisms contribute to diagnosis and discovery in the undiagnosed diseases network: Current state and a future vision. *Orphanet Journal of Rare Diseases*, 16(1), 206. <https://doi.org/10.1186/s13023-021-01839-9>.
- Bello, O. A., Esan, E. B., & Obembe, O. O. (2018). Establishing surface sterilization protocol for nodal culture of *Solanecio biafrae*. *IOP Conference Series: Earth and Environmental Science*, 210(1), 012007. IOP Publishing. <https://doi.org/10.1088/1755-1315/210/1/012007>.
- Berestetskiy, A. (2023). Modern approaches for the development of new herbicides based on natural compounds. *Plants*, 12(2), 234. <https://doi.org/10.3390/plants12020234>.
- Bitwell, C., Indra, S. S., Luke, C., & Kakoma, M. K. (2023). A review of modern and conventional extraction techniques and their applications for extracting phytochemicals from plants. *Scientific African*, 19, e01585. <https://doi.org/10.1016/j.sciaf.2023.e01585>.
- Bourgeois, B., Munoz, F., Fried, G., Mahaut, L., Armengot, L., Denelle, P., Storkey, J., Gaba, S., & Violle, C. (2018). What makes a weed a weed? A large-scale evaluation of arable weeds through a functional lens. *American Journal of Botany*, 106(1), 90-100. <https://doi.org/10.1002/ajb2.1213>.
- Bošnjak Mihovilović, A., Kereša, S., Lazarević, B., Topolovec Pintarić, S., Martinko, K., Marković, Z., Turkalj, K., & Habuš Jerčić, I. (2024). The use of sodium hypochlorite and plant preservative mixture significantly reduces seed-borne pathogen contamination when establishing in vitro cultures of wheat (*Triticum aestivum* L.) seeds. *Agriculture*, 14(4), 556. <https://doi.org/10.3390/agriculture14040556>.
- Budiman, Sandiah, N., & La Malesi. (2020). Growth response of Beha grass (*Brachiaria humidicola*) fertilized with goat manure. *Indonesian Journal of Animal Agricultural Science*, 2(3), 125-127.
- Burdejova, L., Tobolkova, B., Polovka, M., & Neugebauerova, J. (2023). Differentiation of medicinal plants according to solvents, processing, origin, and season by means of multivariate analysis of

- spectroscopic and liquid chromatography data. *Molecules*, 28(10), 4075. <https://doi.org/10.3390/molecules28104075>.
- Cahyo, A. N., Sahuri, I. S. N., & Ardika, R. (2019). Cocopeat as soil substitute media for rubber (*Hevea brasiliensis* Müll. Arg.) planting material. *Journal of Tropical Crop Science*, 6(1), 18-29. <http://dx.doi.org/10.29244/jtcs.6.01.18-29>.
- Cartner, T., Brand, N., Tian, K., Saud, A., Carr, T., Stapleton, P., Lane, M. E., & Rawlings, A. V. (2017). Effect of different alcohols on stratum corneum kallikrein 5 and phospholipase A2 together with epidermal keratinocytes and skin irritation. *International Journal of Cosmetic Science*, 39(2), 188-196. <https://doi.org/10.1111/ics.12364>.
- Casimero, M., Abit, M. J., Ramirez, A. H., Dimaano, N. G., & Mendoza, J. (2023). Herbicide use history and weed management in Southeast Asia. *Advances in Weed Science*, 40, e020220054. <https://doi.org/10.51694/AdvWeedSci/2022;40:seventy-five013>.
- Cesarino, I., Ioio, R. D., Kirschner, G. K., Ogden, M. S., Picard, K. L., Rast-Somssich, M. I., & Somssich, M. (2020). Plant science's next top models. *Annals of Botany*, 126(1), 1. <https://doi.org/10.1093/aob/mcaa063>.
- Cheng, J., Huang, H., Liu, W., Zhou, Y., Han, W., Wang, X., & Zhang, Y. (2022). Unraveling the effects of cold stratification and temperature on the seed germination of invasive *Spartina alterniflora* across latitude. *Frontiers in Plant Science*, 13, 911804. <https://doi.org/10.3389/fpls.2022.911804>.
- Chen, J., Jin, Z., Xiang, L., Chen, Y., Zhang, J., Zhao, J., Huang, F., Shi, Y., Cheng, F., & Pan, G. (2023). Ethanol suppresses rice seed germination through inhibiting ROS signaling. *Journal of Plant Physiology*, 291, 154123. <https://doi.org/10.1016/j.jplph.2023.154123>.
- Chung, I., Ryu, H., Yoon, Y., & Ha, J. C. (2022). Health effects of sodium hypochlorite: Review of published case reports. *Environmental Analysis, Health and Toxicology*, 37(1), e2022006. <https://doi.org/10.5620/eaht.2022006>.
- Cui, Y., He, M., Liu, D., Liu, J., Liu, J., & Yan, D. (2023). Intercellular communication during stomatal development with a focus on the role of symplastic connection. *International Journal of Molecular Sciences*, 24(3), 2593. <https://doi.org/10.3390/ijms24032593>.
- Dadang, Hartono, A., Nurulali, L., & Soekarno, B. P. W. (2019). Effects of paraquat dichloride application on soil arthropods and soil chemical and physical properties in oil palm cultivation. *Journal of International Society for Southeast Asian Agricultural Sciences (J. ISSAAS)*, 25(2), 174-184.
- Daud, Y., & Manu, T. S. (2021). Etnobotani tumbuhan obat oleh masyarakat Desa Pukuafu, Kecamatan Landu Leko, Kabupaten Rote Ndao (Etnobotany of medicine plants by the community of Pukuafu Village, Landu Leko District, Rote Ndao District). *Indigenous Biologi Jurnal Pendidikan dan Sains Biologi*, 4(3).
- Day, M. D., & Callander, J. T. (2024). The benefits and potential of pre-emptive weed biological control: Three case studies in Queensland, Australia. *Biological Control*, 198, 105635. <https://doi.org/10.1016/j.biocontrol.2024.105635>.
- Dewanti, P., & Hariyono, K. (2022). Tobacco Plant (*Nicotiana tabacum* L.) Tolerance towards waterlogging stress on various calcium fertilizer application. *Jurnal Agronomi Indonesia (Indonesian Journal of Agronomy)*, 50(2), 226-233. <https://dx.doi.org/10.24831/jai.v50i2.40431>.

- Dewi, L. K., Setyawati, S. D., Pamuji, A. N. F., Indrayana, S., & Cahyani, C. (2023). The effect of various solvent in Soxhlet extraction on the characteristics of basil oil (*Ocimum americanum* L.). *Jurnal Bahan Alam Terbarukan*, 12(1), 63-69.
- Dias, M. C., Pinto, D. C., & Silva, A. M. (2021). Plant flavonoids: Chemical characteristics and biological activity. *Molecules*, 26(17), 5377. <https://doi.org/10.3390/molecules26175377>.
- Djamal, J. M., Merari, J., & Rukmana, R. M. (2020). Aktivitas sitotoksik dan antiangiogenesis umbi mentimun papasan (*Coccinia grandis* L. Voight) terhadap sel kanker HeLa yang diinduksi protein bFGF. *Riset Informasi Kesehatan*, 9(1), 51-63. <https://doi.org/10.30644/rik.v7i1.218>.
- Duke, S. O., Twitty, A., Baker, C., Sands, D., Boddy, L., Travaini, M. L., ... & Dayan, F. E. (2024). New approaches to herbicide and bioherbicide discovery. *Weed Science*, 72(5), 444-464. <https://doi.org/10.1017/wsc.2024.54>.
- Fatica, A., Di Lucia, F., Marino, S., Alvino, A., Zuin, M., De Feijter, H., Brandt, B., Tommasini, S., Fantuz, F., & Salimei, E. (2019). Study on analytical characteristics of *Nicotiana tabacum* L., cv. *Solaris* biomass for potential uses in nutrition and biomethane production. *Scientific Reports*, 9(1), 1-8. <https://doi.org/10.1038/s41598-019-53237-8>.
- Febryanto, M.A. (2017). Studi ekstraksi dengan metode Soxhletasi pada bahan organik umbi sarang semut (*Myrmecodia pendans*) sebagai inhibitor organik. Institut Teknologi Sepuluh Nopember.
- Ferjani, A., Tsukagoshi, H., & Vassileva, V. (2023). Editorial: Model organisms in plant science: *Arabidopsis thaliana*. *Frontiers in Plant Science*, 14, 1279230. <https://doi.org/10.3389/fpls.2023.1279230>.
- Gaines, T. A., Duke, S. O., Morran, S., G Rigon, C. A., Tranel, P. J., Küpper, A., & Dayan, F. E. (2020). Mechanisms of evolved herbicide resistance. *The Journal of Biological Chemistry*, 295(30), 10307. <https://doi.org/10.1074/jbc.REV120.013572>.
- Ge, Y., Liu, X., Nan, F., Liu, Q., Lv, J., Feng, J., & Xie, S. (2021). Toxicological effects of mercuric chloride exposure on *Scenedesmus quadricauda*. *Water*, 14(20), 3228. <https://doi.org/10.3390/w14203228>.
- Ghadge, A. G., Karmakar, K., Devani, R. S., Banerjee, J., Mohanasundaram, B., Sinha, R. K., ... & Banerjee, A. K. (2014). Flower development, pollen fertility and sex expression analyses of three sexual phenotypes of *Coccinia grandis*. *BMC Plant Biology*, 14, 1-15. <https://doi.org/10.1186/s12870-014-0325-0>.
- Giraldi, R. P., & Rahmadi, R. (2023). Efektivitas herbisida parakuat diklorida dalam mengendalikan gulma perkebunan karet (*Hevea brasiliensis*). *J-Plantasimbiosa*, 5(1), 19-28.
- Gopalsatheeskumar, K. (2018). Significant role of Soxhlet extraction process in phytochemical research. *Mintage Journal of Pharmaceutical & Medical Sciences*, 7(1), 43-47.
- Govindaswamy, V. (2015). Therapeutical properties of ferulic acid and bioavailability enhancement through feruloyl esterase. *Journal of Functional Foods*, 17, 657-666. <https://doi.org/10.1016/j.jff.2015.06.013>.
- Guo, Y., Liu, C., Zhang, Y., Zheng, S., Cao, P., Wang, X., & Tian, Z. (2024). Characterization key genes of *Arabidopsis* seedlings in response to β -caryophyllene, eugenol using combined transcriptome and WGCN analysis. *Frontiers in Plant Science*, 14, 1295779. <https://doi.org/10.3389/fpls.2023.1295779>.

- Hakim, D. B., Hadianto, A., Hutaria, T., & Amaliah, S. (2020). The production efficiency of herbicides in palm oil plantation in Sumatera and Kalimantan. *IOP Conference Series: Earth and Environmental Science*, 468(1), 012054. IOP Publishing. <https://doi.org/10.1088/1755-1315/468/1/012054>.
- Hasan, M., Mokhtar, A. S., Mahmud, K., Rosli, A. M., Hamdan, H., Motmainna, MST., & Ahmad-Hamdani, M. S. (2023). Weed control efficacy and soil activity of a new promising bioherbicide 'WeedLock'. *Sains Malaysiana*, 52(8), 2225-2235. <http://doi.org/10.17576/jsm-2023-5208-05>.
- Heap, I., & Duke, S. O. (2018). Overview of glyphosate-resistant weeds worldwide. *Pest Management Science*, 74(5), 1040-1049. <https://doi.org/10.1002/ps.4760>.
- Hewitt, A. J., Galea, V. J., & O'Donnell, C. (2023). Application technology for bioherbicides: Challenges and opportunities with dry inoculum and liquid spray formulations. *Pest Management Science*, 80(1), 72-80. <https://doi.org/10.1002/ps.7823>.
- He, Y., Galaj, E., Bi, H., Wang, F., Gardner, E., & Xi, X. (2020). β -Caryophyllene, a dietary terpenoid, inhibits nicotine taking and nicotine seeking in rodents. *British Journal of Pharmacology*, 177(9), 2058. <https://doi.org/10.1111/bph.14969>.
- Hornýák, M., Dziurka, M., Pastuszak, J., Szczerba, A., Szklarczyk, M., & Płażek, A. (2022). Photosynthetic efficiency, growth and secondary metabolism of common buckwheat (*Fagopyrum esculentum* Moench) in different controlled-environment production systems. *Scientific Reports*, 12(1), 1-13. <https://doi.org/10.1038/s41598-021-04134-6>.
- Hossain, M. S., Jahan, I., Islam, M., Nayeem, J., Anzum, T. S., Afrin, N. A., Mim, F. K., & Hasan, M. K. (2024). *Coccinia grandis*: Phytochemistry, pharmacology and health benefits. *Clinical Traditional Medicine and Pharmacology*, 5(2), 200150. <https://doi.org/10.1016/j.ctmp.2024.200150>.
- Islam, A. K. M. M., Karim, S. M. R., Kheya, S. A., & Yeasmin, S. (2024). Unlocking the potential of bioherbicides for sustainable and environment friendly weed management. *Heliyon*, 10(16), e36088. <https://doi.org/10.1016/j.heliyon.2024.e36088>.
- Iyer, A., Bestwick, C. S., Duncan, S. H., & Russell, W. R. (2021). Invasive plants are a valuable alternate protein source and can contribute to meeting climate change targets. *Frontiers in Sustainable Food Systems*, 5, 575056. <https://doi.org/10.3389/fsufs.2021.575056>.
- Jayusman, Hakim, L., & Dalimunthe, A. (2022). Season, basal media and plant growth regulators effect in wood plant in vitro propagation: A comprehensive review. *IOP Conference Series: Earth and Environmental Science*, 1115(1), 012051. IOP Publishing. <https://doi.org/10.1088/1755-1315/1115/1/012051>.
- Jia, L., Sun, M., He, M., Yang, M., Zhang, M., & Yu, H. (2024a). Study on the change of global ecological distribution of *Nicotiana tabacum* L. based on MaxEnt model. *Frontiers in Plant Science*, 15, 1371998. <https://doi.org/10.3389/fpls.2024.1371998>.
- Jia, M., Wang, X., Zhu, X., Du, Y., Zhou, P., Wang, G., Wang, N., & Bai, Y. (2024). Accumulation of coumaric acid is a key factor in tobacco continuous cropping obstacles. *Frontiers in Plant Science*, 15, 1477324. <https://doi.org/10.3389/fpls.2024.1477324>.
- Jibrin, M. O., Liu, Q., Jones, J. B., & Zhang, S. (2021). Surfactants in plant disease management: A brief review and case studies. *Plant Pathology*, 70(3), 495-510. <https://doi.org/10.1111/ppa.13318>.

- Joo, N., & Lee, I. (2022). Identification and quantification of key phytochemicals, phytohormones, and antioxidant properties in *Coccinia grandis* during fruit ripening. *Antioxidants*, 11. <https://doi.org/10.3390/antiox11112218>.
- Jugulam, M., & Shyam, C. (2019). Non-target-site resistance to herbicides: Recent developments. *Plants*, 8(10), 417. <https://doi.org/10.3390/plants8100417>.
- Kaab, S., Rebey, I., Hanafi, M., Hammi, K., Smaoui, A., Fauconnier, M., De Clerck, C., Jijakli, M., & Ksouri, R. (2019). Screening of Tunisian plant extracts for herbicidal activity and formulation of a bioherbicide based on *Cynara cardunculus*. *South African Journal of Botany*, 128, 67-76. <https://doi.org/10.1016/j.sajb.2019.10.018>.
- Kacprzyk, J., Schwarze, J., Burke, R., & McCabe, P. (2021). Plant programmed cell death meets auxin signalling. *The FEBS Journal*, 289. <https://doi.org/10.1111/febs.16210>.
- Kamaruddin, N. A., Hanipah, F. A. A., & Zuhari, N. F. (2024). Evaluation of the effect of feeding *Brachiaria humidicola* supplemented with *Azolla* sp. on the growth performance of Saanen doeling (*Capra aegagrus hircus*). *BIO Web of Conferences*, 143, 02001. EDP Sciences. <https://doi.org/10.1051/bioconf/202414302001>.
- Kar, M. M., & Raichaudhuri, A. (2021). Overview of *Arabidopsis* as a genetics model system and its limitation, leading to the development of emerging plant model systems. *Model Organisms in Plant Genetics*. IntechOpen. <http://dx.doi.org/10.5772/intechopen.99818>.
- Kato-Noguchi, H. (2024). Isolation and identification of allelochemicals and their activities and functions. *Journal of Pesticide Science*, 49(1), 1. <https://doi.org/10.1584/jpestics.D23-052>.
- Kekeçoğlu, M., & Sorucu, A. (2021). Determination of the effect of green extraction solvents on the phenolic acids and flavonoids of propolis. *Journal of Research in Veterinary Medicine*, 41(1), 49-54. <https://doi.org/10.30782/jrv.937418>.
- Khamare, Y., Chen, J., & Marble, S. C. (2022). Allelopathy and its application as a weed management tool: A review. *Frontiers in Plant Science*, 13, 1034649. <https://doi.org/10.3389/fpls.2022.1034649>.
- Kondhare, D., & Lade, H. (2017). Phytochemical profile, aldose reductase inhibitory, and antioxidant activities of Indian traditional medicinal *Coccinia grandis* (L.) fruit extract. *3 Biotech*, 7(6), 378. <https://doi.org/10.1007/s13205-017-1013-1>.
- Kostina-Bednarz, M., Płonka, J., & Barchanska, H. (2023). Allelopathy as a source of bioherbicides: Challenges and prospects for sustainable agriculture. *Reviews in Environmental Science and Bio/Technology*, 22(2), 471-504. <https://doi.org/10.1007/s11157-023-09656-1>.
- Kubiak, A., Niewiadomska, A., & Pilarska, A. A. (2022). The problem of weed infestation of agricultural plantations vs. the assumptions of the European biodiversity strategy. *Agronomy*, 12(8), 1808. <https://doi.org/10.3390/agronomy12081808>.
- Kumalasari, N.R., Prasetyowati, A., & Suryahadi. (2021). Estimation of weed biomass as forage production under palmoil plantation. *Jurnal Ilmu Lingkungan*, 21(3), 712-716. <https://doi.org/10.14710/jil.21.3.712-716>.
- Kuppusamy, S., Ramanathan, S., Sengodagounder, S., Senniappan, C., Shanmuganathan, R., Brindhadevi, K., & Kaliannan, T. (2019). Optimizing the sterilization methods for initiation of the five different clones of the *Eucalyptus* hybrid species. *Biocatalysis and Agricultural Biotechnology*, 22, 101361. <https://doi.org/10.1016/j.bcab.2019.101361>.

- Kurniawan, E., Ishak, & Suryani. (2019). Utilization of cocopeat and goat of dirt in marking of solid organic fertilizer to quality macro nutrient (NPK). *IOP Conference Series: Materials Science and Engineering*, 543(1), 012001. IOP Publishing.
- Laboni, F. R., Sultana, T., Kamal, S., Karim, S., Das, S., Harun-Or-Rashid, M., & Shahriar, M. (2017). Biological investigations of the ethanol extract of the aerial part (leaf) of *Coccinia grandis* L. *Journal of Pharmacognosy and Phytochemistry*, 6(2), 134-138.
- Latif, S., Chiapusio, G., & Weston, L. A. (2017). Allelopathy and the role of allelochemicals in plant defence. *Advances in Botanical Research*, 82, 19-54. Academic Press. <http://dx.doi.org/10.1016/bs.abr.2016.12.001>.
- Lee, I. Y., & Joo, N. (2022). Identification and quantification of key phytochemicals, phytohormones, and antioxidant properties in *Coccinia grandis* during fruit ripening. *Antioxidants*, 11(11), 2218. <https://doi.org/10.3390/antiox11112218>.
- Lee, J., Jayakody, J. T., Kim, J., Jeong, J., Choi, K., Kim, T., Seo, C., Azimi, I., Hyun, J., & Ryu, B. (2023). The influence of solvent choice on the extraction of bioactive compounds from Asteraceae: A comparative review. *Foods*, 13(19), 3151. <https://doi.org/10.3390/foods13193151>.
- Lencucha, R., Drope, J., Magati, P., & Sahadewo, G. A. (2022). Tobacco farming: Overcoming an understated impediment to comprehensive tobacco control. *Tobacco Control*, 31(2), 308-312. <https://doi.org/10.1136/tobaccocontrol-2021-056564>.
- Liu, M., Liu, X., Song, Y., Hu, Y., Yang, C., Li, J., Jin, S., Gu, K., Yang, Z., Huang, W., Su, J., & Wang, L. (2024). Tobacco production under global climate change: Combined effects of heat and drought stress and coping strategies. *Frontiers in Plant Science*, 15, 1489993. <https://doi.org/10.3389/fpls.2024.1489993>.
- Lopes, R. W., Marques Morais, E., Lacerda, J. J., & Araújo, F. D. (2022). Bioherbicidal potential of plant species with allelopathic effects on the weed *Bidens bipinnata* L. *Scientific Reports*, 12(1), 1-12. <https://doi.org/10.1038/s41598-022-16203-5>.
- Lorenzo, P., & Morais, M. C. (2024). Repurposing waste from aggressive acacia invaders to promote its management in large invaded areas in Southwestern Europe. *Plants*, 13(11), 1428.
- Luo, Q., Zhang, D., Zhou, J., Qin, M., Ntezimana, B., Jiang, X., Zhu, J., Yu, Z., Chen, Y., & Ni, D. (2024a). Oxidation of tea polyphenols promotes chlorophyll degradation during black tea fermentation. *Food Research International*, 196, 115016. <https://doi.org/10.1016/j.foodres.2024.115016>.
- Luo, Y., Dao, G., Zhou, G., Wang, Z., Xu, Z., Lu, X., & Pan, X. (2024b). Effects of low concentration of gallic acid on the growth and microcystin production of *Microcystis aeruginosa*. *Science of The Total Environment*, 916, 169765. <https://doi.org/10.1016/j.scitotenv.2024.169765>.
- Mahmoud, S. N., & Al-Ani, N. K. (2016). Effect of different sterilization methods on contamination and viability of nodal segments of *Cestrum nocturnum* L. *International Journal of Research Studies in Biosciences*, 4(1), 4-9. <http://dx.doi.org/10.20431/2349-0365.0401002>.
- Matras, E., Gorczyca, A., Pocięcha, E., Przemieniecki, S. W., & Oćwieja, M. (2022). Phytotoxicity of silver nanoparticles with different surface properties on monocots and dicots model plants. *Journal of Soil Science and Plant Nutrition*, 22(2), 1647-1664. <https://doi.org/10.1007/s42729-022-00760-9>.
- Misganaw, G., & Mutai, C. (2024). The contribution of microbial endophytes associated with climate-smart brachiaria grass species to sustainable agriculture and environment. *Land and*

- Water Degradation in Ethiopia: Climate and Land Use Change Implications*, 43-69. Cham: Springer Nature Switzerland. https://doi.org/10.1007/978-3-031-60251-1_4.
- Muhammad, Z. I., Maria, K. S., Mohammad, A., Muhammad, S. H. A. F. I. Q., Zia-Ur-Rehman, F., & Muhammad, K. A. B. I. R. (2015). Effect of mercury on seed germination and seedling growth of Mungbean (*Vigna radiata* (L.) Wilczek). *Journal of Applied Sciences and Environmental Management*, 19(2), 191-199. <http://dx.doi.org/10.4314/jasem.v19i2.4>.
- Muraleedharan, A., Bupesh, G., Vasanth, S., & Balaji, K. (2022). Physicochemical, phytochemical and antioxidant efficacy of the medicinal plant *Coccinia grandis*. *International Journal of Health Sciences*, 6(S4), 7110–7122. <https://doi.org/10.53730/ijhs.v6nS4.10151>.
- Mutai, C., Njuguna, J., & Ghimire, S. (2017). Brachiaria Grasses (*Brachiaria* spp.) harbor a diverse bacterial community with multiple attributes beneficial to plant growth and development. *MicrobiologyOpen*, 6(5), e00497. <https://doi.org/10.1002/mbo3.497>.
- Namazzi, C., Sserumaga, J. P., Mugerwa, S., Kyalo, M., Mutai, C., Mwesigwa, R., Djikeng, A., & Ghimire, S. (2020). Genetic diversity and population structure of *Brachiaria* (syn. *Urochloa*) ecotypes from Uganda. *Agronomy*, 10(8), 1193. <https://doi.org/10.3390/agronomy10081193>.
- Namchaiw, P., Jaisin, Y., Niwaspragrit, C., Malaniyom, K., Auvuchanon, A., & Ratanachamnong, P. (2021). The leaf extract of *Coccinia grandis* (L.) Voigt accelerated in vitro wound healing by reducing oxidative stress injury. *Oxidative Medicine and Cellular Longevity*, 2021(1), 3963510. <https://doi.org/10.1155/2021/3963510>.
- Nayak, K., Tiwari, R., Parte, V., Chouhan, M., Kumar, P., Verma, B., Jayant, R., & Pandey, A. Bio-herbicides: An eco-friendly approach for integrated weed management. *International Journal of Advanced Biochemistry Research*, 8(5), 828-840. <http://dx.doi.org/10.33545/26174693.2024.v8.i5j.1201>.
- Nazish, T., Huang, J., Zhang, J., Xia, Q., Alfatih, A., Luo, C., Cai, T., Xi, J., Xu, P., & Xiang, B. (2022). Understanding paraquat resistance mechanisms in *Arabidopsis thaliana* to facilitate the development of paraquat-resistant crops. *Plant Communications*, 3(3), 100321. <https://doi.org/10.1016/j.xplc.2022.100321>.
- Neamsuvan, O., Komonhiran, P., & Boonming, K. (2018). Medicinal plants used for hypertension treatment by folk healers in Songkhla province, Thailand. *Journal of Ethnopharmacology*, 214, 58-70. <https://doi.org/10.1016/j.jep.2017.11.032>.
- Niazi, P., & Monib, A. W. (2023). Function of macronutrients in plant growth and human. *International Journal of Scientific Development and Research*, 8(4), 1265-1274. <http://dx.doi.org/10.1729/Journal.33883>.
- Niedrite, E., Klavins, L., Dobkevica, L., Purmalis, O., Ievinsh, G., & Klavins, M. (2024). Sustainable control of invasive plants: Compost production, quality and effects on wheat germination. *Journal of Environmental Management*, 371, 123149. <https://doi.org/10.1016/j.jenvman.2024.123149>.
- Nugroho, R., & Setijaningrum, E. (2024). Policy analysis of tobacco excise sharing fund (Rocchipi method). *Indonesian Treasury Review: Jurnal Perbendaharaan, Keuangan Negara dan Kebijakan Publik*, 9(3), 169-185.
- Ocán-Torres, D., Martínez-Burgos, W. J., Manzoki, M. C., Soccol, V. T., Dalmás Neto, C. J., & Soccol, C. R. (2024). Microbial bioherbicides based on cell-free phytotoxic metabolites: Analysis and perspectives on their application in weed control as an innovative sustainable solution. *Plants*, 13(14), 1996. <https://doi.org/10.3390/plants13141996>.

- Pan, X., Zheng, Y., Lei, K., Tao, W., & Zhou, N. (2024). Systematic analysis of Heat Shock Protein 70 (HSP70) gene family in radish and potential roles in stress tolerance. *BMC Plant Biology*, 24(1), 2. <https://doi.org/10.1186/s12870-023-04653-6>
- Panozzo, S., Mascanzoni, E., Scarabel, L., Milani, A., Dalazen, G., Merotto, A. J., Tranel, P. J., & Sattin, M. (2021). Target-site mutations and expression of ALS gene copies vary according to *Echinochloa* species. *Genes*, 12(11), 1841. <https://doi.org/10.3390/genes12111841>.
- Pasternak, T. P., & Steinmacher, D. (2023). Plant growth regulation in cell and tissue culture in vitro. *Plants*, 13(2), 327. <https://doi.org/10.3390/plants13020327>.
- Patil, J. R., Mhatre, K. J., Yadav, K., Yadav, L. S., Srivastava, S., & Nikalje, G. C. (2024). Flavonoids in plant-environment interactions and stress responses. *Discover Plants*, 1(1), 1-19. <https://doi.org/10.1007/s44372-024-00063-6>.
- Peduruhewa, P. S., Jayathunage, K. G. L. R., & Liyanage, R. (2022). Phytochemical screening and antioxidants in vitro bioaccessibility of *Coccinia grandis*: An underutilized wild edible plant in Sri Lanka. *IOP Conference Series: Earth and Environmental Science*, 1094(1), 012007. IOP Publishing. <https://doi.org/10.1088/1755-1315/1094/1/012007>.
- Perico, C., Tan, S., & Langdale, J. A. (2022). Developmental regulation of leaf venation patterns: monocot versus eudicots and the role of auxin. *New Phytologist*, 234(3), 783-803. <https://doi.org/10.1111/nph.17955>.
- Pratyusha, S. (2022). Phenolic compounds in the plant development and defense. *Plant stress physiology: Perspectives in Agriculture*, 125. <https://doi.org/10.5772/intechopen.102873>.
- Prihantoro, I., Aditia, E. L., Setiana, M. A., Saidah, I., & Meilania, R. (2023). Peningkatan produksi rumput *Brachiaria humidicola* pada padang penggembalaan melalui suplementasi pupuk organik feses ayam. *Jurnal Agripet*, 23(2), 196-204. <https://doi.org/10.17969/agripet.v23i2.27848>.
- Puhili, A. L., Chikmawati, T., & Rugayah, R. (2022). *Coccinea grandis* (L.) Voigt.(Cucurbitaceae): Morfologi dan persebarannya di Papua. *Floribunda*, 7(1), 26-29.
- Putra, I. M. W. A., Kusumawati, I. G. A. W., & Sumadewi, N. L. U. (2021). Physical characteristics, total phenolic, and flavonoid content of *Coccinia grandis* (L.) Voigt leaves extract. *Acta Chimica Asiana*, 4(2), 114-119. <https://doi.org/10.29303/aca.v4i2.66>.
- Putri, K. D., Guntoro, D., & Ardie, S. W. (2024). A new amino acid substitution in the MvALS1 gene of metsulfuron-methyl resistant biotypes *Monochoria vaginalis* (Burm. F.) C. Presl from West Java, Indonesia. *PLOS ONE*, 19(10), e0308465. <https://doi.org/10.1371/journal.pone.0308465>.
- Ramandi, A., Javan, I. Y., Tazehabadi, F. M., Asl, G. I., Khosravanian, R., & Ebrahimzadeh, M. H. (2019). Improvement in seed surface sterilization and in vitro seed germination of ornamental and medicinal plant-*Catharanthus roseus* (L.). *Chiang Mai Journal of Science*, 46(6), 1107-1112.
- Rasouli, H., Farzaei, M. H., Mansouri, K., Mohammadzadeh, S., & Khodarahmi, R. (2016). Plant cell cancer: May natural phenolic compounds prevent onset and development of plant cell malignancy? A literature review. *Molecules*, 21(9), 1104. <https://doi.org/10.3390/molecules21091104>.
- Roberts, J., Florentine, S., Fernando, W. G., & Tennakoon, K. U. (2021). Achievements, developments and future challenges in the field of bioherbicides for weed control: A global review. *Plants*, 11(17), 2242. <https://doi.org/10.3390/plants11172242>.

- Roy, A., Khan, A., Ahmad, I., Alghamdi, S., Rajab, B. S., Babalghith, A. O., Alshahrani, M. Y., Islam, S., & Islam, M. R. (2022). Flavonoids a bioactive compound from medicinal plants and its therapeutic applications. *BioMed Research International*, 2022, 5445291. <https://doi.org/10.1155/2022/5445291>.
- Rusdy, M., Sjahril, R., Riadi, M., & Budiman, B. (2013). Integration of mechanical and cultural control treatments to manage invasive shrub *Chromolaena odorata* and other weeds under drought conditions in pasture area. *Journal of the Indonesian Tropical Animal Agriculture*, 38(1), 65-71.
- Sahari, B., Hendarjanti, H., Yusran, A., Ibrahim, M. I. M., Ramadhan, G. F., & Prabowo, R. (2023). Weed diversity in oil palm plantation: Benefit from the unexpected ground cover community. *IOP Conference Series: Earth and Environmental Science*, 1220(1), 012011. IOP Publishing.
- Sahu, P. K., Tilgam, J., Mishra, S., Hamid, S., Gupta, A., Jayalakshmi, K., Verma, S. K., & Kharwar, R. N. (2022). Surface sterilization for isolation of endophytes: Ensuring what (not) to grow. *Journal of Basic Microbiology*, 62(6), 647-668. <https://doi.org/10.1002/jobm.202100462>.
- Sakharkar, P., & Chauhan, B. (2017). Antibacterial, antioxidant and cell proliferative properties of *Coccinia grandis* fruits. *Avicenna Journal of Phytomedicine*, 7(4), 295. <https://pmc.ncbi.nlm.nih.gov/articles/PMC5580868/>.
- Santoso, A. D., & Purnomo, P. (2021). Variation and phenetic relationship of tobacco (*Nicotiana tabacum* L.) in Central Java and Yogyakarta based on morphological characters. *Jurnal Riset Biologi dan Aplikasinya*, 3(2), 73-79.
- Satriawan, H., & Fuady, Z. (2019). Analysis of weed vegetation in immature and mature oil palm plantations. *Biodiversitas Journal of Biological Diversity*, 20(11). <https://doi.org/10.13057/biodiv/d201123>.
- Scavo, A., & Mauromicale, G. (2021). Crop allelopathy for sustainable weed management in agroecosystems: Knowing the present with a view to the future. *Agronomy*, 11(11), 2104. <https://doi.org/10.3390/agronomy11112104>.
- Selwal, N., Tabassum, Z., Rahayu, F., Yulia, N. D., Sugiono, S., Endarto, O., Rijaya, P. D., Djajadi, D., Khamidah, A., & Wani, A. K. (2023). Therapeutic potential and phytoremediation capabilities of the tobacco plant: Advancements through genetic engineering and cultivation techniques. *Biocatalysis and Agricultural Biotechnology*, 52, 102845. <https://doi.org/10.1016/j.bcab.2023.102845>.
- Setyawardhani, D. A., Majid, D. A., & Plawi, R. S. S. (2024). Solid-liquid equilibrium study of binary system saturated fatty acid in short chain alcohols. *Equilibrium Journal of Chemical Engineering*, 8(1), 8-18. <https://dx.doi.org/10.20961/equilibrium.v8i1.80577>.
- Setyawati, T., Narulita, S., Bahri, I.P., & Rahardjo, G.T. (2015). *A guide book to invasive alien plant species in Indonesia*. Research, Development and Innovation Agency. Ministry of Environment and Forestry. https://ksdae.menlhk.go.id/assets/publikasi/A_Guide_Book_of_Invasive_Plant_Species_in_Indonesia.pdf.
- Shah, A., & Smith, D. L. (2020). Flavonoids in agriculture: Chemistry and roles in, biotic and abiotic stress responses, and microbial associations. *Agronomy*, 10(8), 1209. <https://doi.org/10.3390/agronomy10081209>.

- Shen, N., Wang, T., Gan, Q., Liu, S., Wang, L., & Jin, B. (2022). Plant flavonoids: Classification, distribution, biosynthesis, and antioxidant activity. *Food Chemistry*, 383, 132531. <https://doi.org/10.1016/j.foodchem.2022.132531>
- Si, Y., Haxim, Y., & Wang, L. (2022). Optimum sterilization method for in vitro cultivation of dimorphic seeds of the succulent halophyte *Suaeda aralocaspica*. *Horticulturae*, 8(4), 289. <https://doi.org/10.3390/horticulturae8040289>.
- Steve, C. (2016). *Invasive plant risk assessment: Coccinia grandis*. State of Queensland. https://www.daf.qld.gov.au/__data/assets/pdf_file/0009/68616/IPA-Ivy-Gourd-Risk_Assessment.pdf.
- Su, X., Yue, X., Kong, M., Xie, Z., Yan, J., Ma, W., Wang, Y., Zhao, J., Zhang, X., & Liu, M. (2023). Leaf color classification and expression analysis of photosynthesis-related genes in inbred lines of Chinese cabbage displaying minor variations in dark-green leaves. *Plants*, 12(11), 2124. <https://doi.org/10.3390/plants12112124>.
- Sukhoverkov, K. V., & Mylne, J. S. (2021). A systematic approach for finding herbicide synergies. *BioRxiv*, 2021-02. <https://doi.org/10.1101/2021.02.08.430187>.
- Sumaryono, Muslihatin, W., & Ratnadewi, D. (2012). Effect of carbohydrate source on growth and performance of *in vitro* sago palm (*Metroxylon sagu* Rottb.) plantlets. *HAYATI Journal of Biosciences*, 19(2), 88-92. <https://doi.org/10.4308/hjb.19.2.88>.
- Sumekar, Y., Riswandi, D., Widayat, D., & Umiyati, U. (2021). The effect of paraquat dichloride herbicide for weed control in immature oil palm plantations. *International Journal of Multidisciplinary Research and Growth Evaluation*, 2(1), 248-251.
- Sun, M., Mao, Q., Peng, C., Ai, Q., Yang, Y., Tang, K., Liu, F., & Li, H. (2024). Quercetin inhibits hydrogen peroxide-induced cleavage of heat shock protein 90 to prevent glutathione peroxidase 4 degradation via chaperone-mediated autophagy. *Phytomedicine: International Journal of Phytotherapy and Phytopharmacology*, 136, 156286. <https://doi.org/10.1016/j.phymed.2024.156286>.
- Suprihanti, A. (2022, April). Response analysis of tobacco supply in Indonesia. *IOP Conference Series: Earth and Environmental Science*, 1018(1), 012042. IOP Publishing. <https://doi.org/10.1088/1755-1315/1018/1/012042>.
- Suseno, N., Isnaini, N., Suwignyo, B., Suhartanto, B., & Umami, N. (2021). Callus induction and plant regeneration of Brachiaria grass from immature inflorescence explants. *Buletin Peternakan*, 45(2), 84-89. <https://doi.org/10.21059/buletinpeternak.v45i2.43461>.
- Susilawati, Agustina, R. A., Abdi, F., Hafizianor, Trisnu, S., & Seransyah, R. G. (2023). Ground cover plants diversity in Mandiingin hill forest area with a special purpose of South Kalimantan Province, Indonesia. *Russian Journal of Agricultural and Socio-Economic Sciences*, 134(2), 159-164.
- Sutomo, S., Lestari, H. D., Arnida, A., & Sriyono, A. (2019). Simplicia and extracts standardization from Jualing leaves (*Micromelum minutum* Wight & Arn.) from South Kalimantan. *Borneo Journal of Pharmacy*, 2(2), 55-62. <https://doi.org/10.33084/bjop.v2i2.898>.
- Taalat, K., Javed, M. A., Huyop, F. Z., & Kaya, Y. (2021). Plant tissue culture of *Nicotiana tabacum* cv. TAPM 26 and its minimum inhibition against herbicide-Dalapon. *MANAS Journal of Engineering*, 9(Special 1), 35-42. <https://doi.org/10.51354/mjen.839516>.

- Taurina, W., & Andrie, M. (2022). Standardization of simplicia golden sea cucumber (*Stichopus hermanii*) from Pelapis Island, West Kalimantan. *Majalah Obat Tradisional*, 27(2), 146-152.
- Tian, Y., Liu, X., Fan, C., Li, T., Qin, H., Li, X., Chen, K., Zheng, Y., Chen, F., & Xu, Y. (2021). Enhancement of tobacco (*Nicotiana tabacum* L.) seed lipid content for biodiesel production by CRISPR-Cas9-mediated knockout of NtAn1. *Frontiers in Plant Science*, 11, 599474. <https://doi.org/10.3389/fpls.2020.599474>.
- Tong, Z., Huang, Y., Zhu, Q., Fan, L., Xiao, B., & Shen, E. (2024). Retrospect and prospect of *Nicotiana tabacum* genome sequencing. *Frontiers in Plant Science*, 15, 1474658. <https://doi.org/10.3389/fpls.2024.1474658>.
- Torra, J., & Alcantara-De la Cruz, R. (2022). Molecular mechanisms of herbicide resistance in weeds. *Genes*, 13(11), 2025. <https://doi.org/10.3390/genes13112025>.
- Torres-Sánchez, J., Mesas-Carrascosa, F. J., Jiménez-Brenes, F. M., de Castro, A. I., & López-Granados, F. (2021). Early detection of broad-leaved and grass weeds in wide row crops using artificial neural networks and UAV imagery. *Agronomy*, 11(4), 749. <https://doi.org/10.3390/agronomy11040749>
- Utomo, A. R., & Yunus, A. (2021). The effect of Murashige and Skoog (MS) and Growmore fertilizer media composition on growth of Ambon banana plants in vitro. *IOP Conference Series: Earth and Environmental Science*, 637(1), 012013. IOP Publishing. <https://doi.org/10.1088/1755-1315/637/1/012013>.
- Vailina Dsouza, Puja, Rithika, R., & Malathi, R. (2024). Comparative analysis of cocopeat and soil on plant constituents. *Journal of Chemical Health Risks*, 14(3), 2611-2622.
- Van Loi, B. (2024). Effects of planting densities and regenerations on the growth and yield of *Brachiaria humidicola* grown in Thua Thien Hue province. *Hue University Journal of Science: Natural Science*, 133(1B), 13-22.
- Vollmannová, A., Bojňanská, T., Musilová, J., Lidiková, J., & Cifrová, M. (2024). Quercetin as one of the most abundant represented biological valuable plant components with remarkable chemoprotective effects - A review. *Heliyon*, 10(12), e33342. <https://doi.org/10.1016/j.heliyon.2024.e33342>.
- Wahyusi, K. N., Nurmawati, A., & Zaen, Y. (2023). Mass transfer coefficient study of cherry (*Muntingia calabura* L.) leaves alkaloids extraction using ethanol. *International Journal of Eco-Innovation in Science and Engineering (IJEISE)*, 4(2).
- Wang, L., Yu, L., Yin, W., Li, Y., Liu, L., Song, L., & Zhou, Y. (2021). Comparison of different solvents for extraction of oils from by-products of shrimps *Penaeus vannamei* and *Procambarus clarkia*. *Journal of Food Processing and Preservation*, 45(9), e15754. <https://doi.org/10.1111/jfpp.15754>.
- Wang, J., Song, L., Gong, X., Xu, J., & Li, M. (2020). Functions of jasmonic acid in plant regulation and response to abiotic stress. *International Journal of Molecular Sciences*, 21(4), 1446. <https://doi.org/10.3390/ijms21041446>
- Wardhani, I. Y., Ramadani, A. H., Widyaningrum, F., & Famelia, V. (2023). The effect of extraction method on total flavonoid content of *Ageratum conyzoides* ethanol extract. *Journal of Biology Education*, 6(2), 134-145.
- Warfa'ni, I., Prayoga, L., Prasetyo, R., Murchie, E. H., & Sugiyono. The effect of media types and NAA concentrations on agarwood (*Aquilaria malaccensis* Lamk.) shoot development in *in vitro* culture.

- AgriHealth: Journal of Agri-food, Nutrition and Public Health*, 3(1), 62-71. <http://dx.doi.org/10.20961/agrihealth.v3i1.60346>.
- Winarti, S., & Kristina. (2018). Growth and yield of grass *Brachiaria humidicola* given chicken manure and NPK fertilizer on sandy soil. *Jurnal Agripeat*, 19(1), 15-22.
- Wu, Y., Ren, D., Gao, C., Li, J., Du, B., Wang, Z., & Qian, S. (2021). Recent advances for alkaloids as botanical pesticides for use in organic agriculture. *International Journal of Pest Management*, 69(3), 288-298. <https://doi.org/10.1080/09670874.2021.1917723>.
- Xie, L., Liu, D., Müller, C., Jansen-Willems, A., Chen, Z., Niu, Y., Zaman, M., Meng, L., & Ding, W. (2022). *Brachiaria humidicola* cultivation enhances soil nitrous oxide emissions from tropical grassland by promoting the denitrification potential: A ¹⁵N tracing study. *Agriculture*, 12(11), 1940. <https://doi.org/10.3390/agriculture12111940>.
- Xu, L., & Wang, X. (2025). A Comprehensive Review of Phenolic Compounds in Horticultural Plants. *International Journal of Molecular Sciences*, 26(12), 5767. <https://doi.org/10.3390/ijms26125767>.
- Yadav, L. P., Gangadhara, K., Apparao, V. V., Yadav, V., Mishra, D. S., Singh, A. K., Rane, J., Kaushik, P., Janani, P., Kumar, R., Verma, A. K., Kumar, S., Malhotra, S. K., & Shekhawat, N. (2024). Genetic diversity, morphological traits, quality traits and antioxidants potentiality of *Coccinia grandis* germplasm under rainfed semi-arid region. *Scientific Reports*, 14(1), 1-17. <https://doi.org/10.1038/s41598-023-49091-4>.
- Yalcin, H., Kavuncuoğlu, H., Tulukcu, E., & Eroğlu, Z. (2017). The effect of harvest time on the bioactive properties and volatile components of lavender (*Lavandula officinalis*). *Quality Assurance and Safety of Crops & Foods*, 9(3), 275-283. <http://dx.doi.org/10.3920/QAS2015.0763>.
- Yang, L. Y., Yang, S. L., Li, J. Y., Ma, J. H., Pang, T., Zou, C. M., He, B., & Gong, M. (2018). Effects of different growth temperatures on growth, development, and plastid pigments metabolism of tobacco (*Nicotiana tabacum* L.) plants. *Botanical Studies*, 59(1), 1-13. <https://doi.org/10.1186/s40529-018-0221-2>.
- Yiamthaisong, N., Jampeetong, A., & Tiansawat, P. (2024). Surface sterilization and moist storage conditions for recalcitrant seeds of three Asian tree species. *Forest Science and Technology*, 20(4), 410-417. <https://doi.org/10.1080/21580103.2024.2409916>.
- Zahara, M., Datta, A., Boonkorkaew, P., & Mishra, A. (2017). The effects of different media, sucrose concentrations and natural additives on plantlet growth of *Phalaenopsis* hybrid 'Pink'. *Brazilian Archives of Biology and Technology*, 60. <https://doi.org/10.1590/1678-4324-2017160149>.
- Zhang, Q., Li, M., Yang, G., Liu, X., Yu, Z., & Peng, S. (2022). Protocatechuic acid, ferulic acid and relevant defense enzymes correlate closely with walnut resistance to *Xanthomonas arboricola* pv. *juglandis*. *BMC Plant Biology*, 22(1), 598. <https://doi.org/10.1186/s12870-022-03997-9>.
- Zhang, W., Lin, G., & Ye, C. (2018). Techniques for extraction and isolation of natural products: A comprehensive review. *Chinese Medicine*, 13, 20. <https://doi.org/10.1186/s13020-018-0177-x>.
- Zhang, W., Pan, X., Fu, J., Cheng, W., Lin, H., Zhang, W., & Huang, Z. (2024). Phytochemicals derived from *Nicotiana tabacum* L. plant contribute to pharmaceutical development. *Frontiers in Pharmacology*, 15, 1372456. <https://doi.org/10.3389/fphar.2024.1372456>.
- Zhang, X., Huang, X., Li, Y., Tao, F., Zhao, Q., & Li, W. (2021). Polar auxin transport may be responsive to specific features of flavonoid structure. *Phytochemistry*, 185, 112702. <https://doi.org/10.1016/j.phytochem.2021.112702>.

- Zhao, N., Yan, Y., Du, L., Zhang, X., Liu, W., & Wang, J. (2020). Unravelling the effect of two herbicide resistance mutations on acetolactate synthase kinetics and growth traits. *Journal of Experimental Botany*, 71(12), 3535-3542. <https://doi.org/10.1093/jxb/eraa120>.
- Zulfisa, Z., Fika, R., Agusfina, M., Yonrizon, Y., & Muhsanah, A. (2023). Determination of total phenolic content of ethanol extract of broken bone twigs (*Euphorbia tirucalli* Linn.) by Folin-Ciocalteu Method Spectrophotometrically. *Jurnal eduhealth*, 14(03), 1326-1331.
- Zumsteg, J., Heitz, T., Heintz, D., Smirnova, E., Marquis, V., Villette, C., Graindorge, S., & Delcros, P. (2021). Broad-spectrum stress tolerance conferred by suppressing jasmonate signaling attenuation in Arabidopsis JASMONIC ACID OXIDASE (JAO) mutants. *The Plant Journal: For Cell and Molecular Biology*. <https://doi.org/10.1111/tpj.15598>.